

Review

Biological Markers in DCIS and Risk of Breast Recurrence: A Systematic Review

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Abstract

Understanding of the biology and clinical behavior of ductal carcinoma in situ (DCIS) is currently inadequate. The aim of this comprehensive review was to identify important molecular biological markers associated with DCIS and candidate markers associated with increased risk of ipsilateral recurrence after diagnosis of DCIS. A comprehensive systematic review was performed to identify studies published in the past 10 years that investigated biological markers in DCIS. To be included in this review, studies that investigated the rate of biological expression of markers had to report on at least 30 patients; studies that analyzed the recurrence risk associated with biomarker expression had to report on at least 50 patients. There were 6,252 patients altogether in our review. Biological markers evaluated included steroid receptors, proliferation markers, cell cycle regulation and apoptotic markers, angiogenesis-related proteins, epidermal growth factor receptor family receptors, extracellular matrix-related proteins, and COX-2. Although the studies in this review provide valuable preliminary information regarding the expression and prognostic significance of biomarkers in DCIS, common limitations of published studies (case-series, cohort, and case-control studies) were that they were limited to small patient cohorts in which the extent of surgery and use of radiotherapy or endocrine therapy varied from patient to patient, and variable methods of determining biomarker expression. These constraints made it difficult to interpret the absolute effect of expression of various biomarkers on risk of local recurrence. No prospective validation studies were identified. As the study of biomarkers are in their relative infancy in DCIS compared with invasive breast cancer, key significant prognostic and predictive markers associated with invasive breast cancer have not been adequately studied in DCIS. There is a critical need for prospective analyses of novel and other known breast cancer molecular markers in large cohorts of patient with DCIS to differentiate indolent from aggressive DCIS and better tailor the need and extent of current therapies.

Key words: Biological Markers, Ductal carcinoma in situ, DCIS, Breast Recurrence

INTRODUCTION

A diagnosis of ductal carcinoma in situ (DCIS) of the breast was rare before the 1980s; however, with the introduction of mammography, the incidence of DCIS grew dramatically. Currently, more than 60,000

patients are diagnosed with DCIS in the United States each year. For many years, mastectomy was the standard procedure to treat DCIS; however, since DCIS is a noninvasive and silent disease,

breast-conserving therapy with or without radiotherapy is now considered sufficient and the accepted method to treat DCIS. A definitive goal in treating DCIS is to prevent ipsilateral invasive breast cancer recurrence as the mortality associated with a diagnosis of pure DCIS is less than 1–2% [1]. About 5–30% of patients with DCIS treated with breast-conserving therapy with or without radiation experience an ipsilateral local recurrence, and about half of these recurrences are invasive. Clinicians are unable to predict the risk of local recurrence or progression to invasive breast cancer in patients with DCIS following their treatment, a major concern that needs to be addressed. Numerous studies have been conducted to address this concern, and in a few studies, investigators have concluded that several histopathologic characteristics of DCIS—lesion size, margin status, nuclear grade, architectural pattern, and presence of necrosis—are predictors of recurrence [1].

The U.S. National Cancer Institute has recently identified a critical need for investigation and validation of molecular factors to improve risk stratification of patients with DCIS, which will facilitate identification of the optimal therapy for each individual patient [2].

A molecular biological marker (“biomarker”) is a molecular feature that is objectively measured and that serves as an indicator of a normal biological response, a pathogenic process, or the likelihood of response to an existing or novel pharmacologic therapy. In cancer, biomarkers may be used for disease detection, disease staging, monitoring of disease response to therapy, and prediction of patient prognosis [3]. A biomarker may be DNA or RNA based or a protein marker measured directly in tissues, serum, or other body fluids [3].

For DCIS, the optimal biomarker or combination of measured biomarkers would provide additional prognostic information beyond that provided by other clinical and pathologic factors established in the field as risk factors for local recurrence, such as lesion grade, lesion size, patient age, margin status, and use versus nonuse of adjuvant radiotherapy. The perfect biomarker or combination of biomarkers and/or clinical pathologic factors would establish which cases of DCIS were highly unlikely to ever be associated with progression to invasive breast cancer and thus which patients needed no further therapy. Biomarker expression might also be useful in individualizing therapy for DCIS with respect to the need for local excision versus mastectomy or the need for adjuvant radiotherapy.

Validation of cancer biomarkers is complex and must be thorough if the marker is to be used in patient

care [4–6]. The aim of this comprehensive review was to identify important biomarkers associated with DCIS and candidate biomarkers associated with increased risk of ipsilateral recurrence after diagnosis of DCIS.

METHODS

A comprehensive search of the MEDLINE and Cochrane databases was performed on February 4, 2011. Search terms included the U.S. National Library of Medicine’s Medical Subject Headings *ductal carcinoma in situ*, *in situ breast cancer*, *intraductal carcinoma*, *stage 0 breast cancer*, and *DCIS*. Additional search terms included *estrogen receptor*, *progesterone receptor*, *androgen receptor*, *Ki-67*, *cyclin D1*, *cyclin A*, *cyclin E*, *p16*, *p21*, *p27*, *p53*, *Bcl-2*, *Bax*, *survivin*, *c-myc*, *Rb*, *VEGF*, *HPR1*, *HER1*, *HER2*, *HER3*, *HER4*, *CD10*, *SPARC* and *COX-2* (Table 1). A total of 622 studies were identified on biological markers in DCIS published in November 2000 or later. One additional significant study published in 1998 and referenced in other studies included in the review was also included [7]. The following types of studies were excluded: in vitro studies; studies involving only animals; case reports; and studies reporting on patients with concurrent invasive breast cancer or non-breast malignancies. Studies that investigated the rate of biological expression of markers had to report on at least 30 patients, and studies that analyzed the recurrence risk associated with biomarker expression had to report on at least 50 patients. There were 6,252 patients altogether in our review.

RESULTS

Biomarker expression rates for different categories of biomarkers and notes on how expression rates were derived are presented in Tables 2–8. Information on the relationship between biomarker expression and the risk of ipsilateral recurrence following surgery for DCIS is presented in Table 9. Table 9 lists the year published, the number of patients in the study, the type of surgery performed, whether post-lumpectomy radiotherapy was given, the median follow-up time, whether endocrine therapy was utilized, and a synopsis of whether the biomarker studied was associated with an increased risk of local recurrence.

Steroid Receptors

Estrogen Receptor

Estrogen receptor (ER) was one of the first biological markers to be studied in depth, and today ER is one of the most valuable markers in breast cancer. ER status in patients with breast cancer predicts re-

sponse to endocrine therapy. Among the 36 studies in our comprehensive review that examined ER expression rate in DCIS, the mean ER expression rate was 68.7% (range: 49–96.6%, Table 2).

Numerous studies revealed ER expression to be inversely related to the nuclear grade of DCIS: ER expression was higher in well-differentiated lesions than in poorly differentiated lesions [8-15]. One study revealed that ER expression was more common in cribriform DCIS than in other DCIS subtypes [7]. Similarly, another study revealed that ER expression was more common in papillary, solid, and cribriform DCIS than in micropapillary and comedocarcinoma DCIS [9].

Many investigators studied the relationship between ER and other biological markers. Several studies revealed ER to be inversely correlated with HER-2/*neu* (HER2) expression [7, 9, 13, 16-18]. One study revealed that ER expression was inversely correlated with p53 expression [7]. A few studies revealed ER expression to be positively associated with high Bcl-2 expression [7, 12, 16]. One study reported a significant correlation between ER negativity and cyclooxygenase-2 (COX-2) positivity [19]; however, another study did not find a significant correlation between ER and COX-2 [20]. Kulkarni et al reported several significant associations between ER and other markers—for instance, progesterone receptor (PR), Cyclin D1, p53, and p21 [21]. Altintas et al reported that lesions with high proliferative activity were more likely to be ER negative [15].

ER Expression and Risk of Local Recurrence

We identified 16 studies (2,470 total patients) that evaluated the relationship between ER expression and risk of local recurrence (Table 9). Four of these studies revealed an association between ER-negative DCIS and risk of local recurrence [16, 22-24]. In the first study, a nested case-control study by Provenzano et al, the investigators compared 53 patients with DCIS who had a subsequent local recurrence to 42 age- and date- of-diagnosis-matched control subjects with DCIS who did not have a recurrence [22]. Patients in the series had breast-conserving surgery with (n=10) or without adjuvant radiotherapy (n=85). Patients with subsequent local recurrence were more likely than those without recurrence to have ER-negative disease (62% vs. 35%; OR: 0.2; $P=0.01$, Table 9) [22].

In the second study that found a relationship between ER-negative DCIS and risk of local recurrence, another nested case-control study, Kerlikowske et al evaluated biomarker expression and risk of local recurrence among 329 patients with DCIS who were

all treated with wide local excision without subsequent radiotherapy. The authors selected tissues to be studied for biomarker expression from 72 women who had a subsequent invasive cancer event, 71 women who had a subsequent DCIS event, and 186 randomly selected women who served as control subjects who did not have a subsequent tumor event [24]. This nested case-control study had a median follow-up time of 8.2 years. According to the univariate analysis, patients with a subsequent DCIS event were more likely than those without a subsequent tumor event to have ER-negative disease (31% vs. 20%, Table 9). ER-negative status was individually associated with DCIS recurrence. In addition, ER-negative status combined with either HER2 expression or Ki-67 expression was associated with DCIS recurrence. Similar results were observed in the multivariate analysis. Patients with DCIS recurrence were more likely to exhibit the phenotype ER-HER2+Ki-67+, and this phenotype was a predictor of DCIS recurrence (hazard ratio: 5.8; 95% confidence interval [CI]: 2.4-14, Table 9). In patients who developed a subsequent *invasive* tumor, the investigators did not find ER to be a predictor of recurrence either individually or in combination with other markers in a phenotype [24]. This study by Kerlikowske et al is one of the largest and arguably one of the most important studies in the field. This study by Kerlikowske et al is valuable because it allows clinicians to study a group of patients who all underwent the identical therapy for DCIS - in this case local excision alone without radiotherapy. Overall, 28% of the patients had a local-regional recurrence. This high rate of recurrence may be related to the fact that 45-61% of patients in the study had positive margins or uncertain margin status [24]. Obtaining definitive negative margins is now a well-known, accepted prerequisite for breast-conserving therapy [1], and this situation illustrates the complexity of interpreting results and the potential clinical value of these types of biomarker studies for present day therapy of DCIS.

In the third study that found a relationship between ER-negative DCIS and risk of local recurrence, Roka et al evaluated 132 patients with DCIS treated with breast-conserving surgery without (n=33) or with whole-breast radiotherapy (n=99) and found that patients with ER-negative DCIS were more likely than those with ER-positive DCIS to have a local recurrence (12.2% vs. 3.7%, Table 9) [23]. This study provides clinically useful information in that it gives an absolute difference between the rates of local recurrence in patients with ER-negative and ER-positive DCIS treated with surgery and radiotherapy: about 8.5%.

Ringberg et al investigated 187 patients with DCIS who underwent breast-conserving therapy with (n=66) or without (n=121) radiotherapy. In the group of patients who did not receive radiotherapy, 31 patients had a recurrence. The authors used principal components analysis to evaluate a cell biological index that included ER and PR negativity, overexpression of HER2, low Bcl-2 expression, accumulation of p53, nondiploidy, and high Ki-67 expression. According to the multivariate analysis, the combination of ER-negative DCIS and the biological markers in the index was a strong predictor of local-regional recurrence (relative risk: 1.3; 95% CI: 1.0-1.6, Table 9) [16].

Zhou et al [25] examined the relationship between triple-negative DCIS (negative for ER, PR, and HER2; also known as *basal-like breast cancer*) and risk of local recurrence. This study included 392 patients, approximately half of whom received adjuvant radiotherapy. Of the 392 patients, 42 patients had an *in situ* recurrence, and 34 had an *invasive* recurrence. The authors compared the 32 patients with triple-negative DCIS with the 360 patients with other phenotypes and found that patients with basal-like DCIS had a higher risk of local recurrence. However, this difference did not reach statistical significance. This study illustrates additional barriers to determining the effect of biomarker expression on local recurrence, including small number of patients and inconsistent use of adjuvant treatments (e.g., radiotherapy), which can confound results.

ER Expression and Response to Therapy

A predictive biomarker is a marker that can predict the response to a specific therapy. For invasive breast cancer, the presence of ER expression and the degree of ER expression have unequivocally been shown to predict response to tamoxifen, which blocks ER, and aromatase inhibitors, which prevent production of estrogen. In women with ER expression, ER has been demonstrated to affect the expression of other relevant biomarkers and lead to increased survival and decreased recurrence risk in women taking tamoxifen or aromatase inhibitors compared to women not taking these drugs [26]. In contrast, no demonstrative effect of tamoxifen or aromatase inhibitors is seen in patients whose invasive tumors lack ER expression.

For DCIS, we found four clinical trials that may shed light on the value of ER for predicting response to tamoxifen and aromatase inhibitors. The first two studies were landmark phase III trials in which patients with DCIS were randomized to tamoxifen or placebo after local excision of DCIS. The first, the UK/ANZ DCIS trial, involved 1,701 patients and had

an unusual randomization scheme involving the use of adjuvant tamoxifen and radiotherapy with elective decision to withhold or provide one of these treatments [27]. The median follow-up time was 12.7 years. Although tamoxifen significantly reduced the incidence of all new breast events ($P<0.0001$), the absolute differences between rates of new breast cancer events in women randomized to tamoxifen and no tamoxifen were small. Patients randomized to tamoxifen had a 3.5% absolute 10-year reduction in the risk of ipsilateral DCIS recurrence, no change in the risk of ipsilateral invasive breast cancer, and a 2.3% absolute 10-year reduction in the risk of new contralateral breast cancer.

The second landmark phase III trial was the National Surgical Adjuvant Breast and Bowel Project (NSABP) B-24 clinical trial, which included 1,804 women, and in which tamoxifen was also investigated among patients receiving radiotherapy [28]. At a median follow-up time of 12 years, there were very small but statistically significant absolute reductions in rates of breast cancer events among patients randomized to tamoxifen versus placebo (a 2.4% reduction in the risk of ipsilateral invasive breast cancer recurrence, no change in the risk of ipsilateral DCIS recurrence, and a 3.2% reduction in the risk of contralateral breast cancer events). An important caveat in interpreting these data is that in both the UK/ANZ and NSABP B-24 trials, patients were not selected to receive tamoxifen on the basis of ER positivity. The magnitude of benefit might have been higher if only patients with ER-positive DCIS had been enrolled. Nevertheless, taken together, the results of these two trials suggest that ER appears to be a weakly predictive biomarker for effecting local recurrence following treatment for DCIS.

The third pivotal trial implicating ER as a predictive biomarker for endocrine manipulation of ER-positive DCIS was reported by Chen et al [29]. This small study, involving just 23 patients, was undertaken to investigate whether neoadjuvant (administered before surgery) endocrine therapy resulted in detectable histologic alterations. In order to determine whether such treatment results in detectable histologic alterations, the authors compared the pathologic and biomarker changes in DCIS following neoadjuvant endocrine therapy (that is, before surgery) to a group of patients who did not undergo preoperative anti-estrogenic treatment. Premenopausal women were treated with tamoxifen, and postmenopausal women were treated with letrozole. Pathologic markers of proliferation, inflammation, and apoptosis were evaluated at baseline and at 3 months. Following treatment, predominant morphologic changes in-

cluded increased multinucleated histiocytes and degenerated cells, a decrease in expression of the proliferation marker Ki-67, and immune infiltration into the lesions consistent with a treatment effect. This study was significant because of its novel clinical trial design, the so-called "window study design," which allows investigators to test potentially promising agents for DCIS and immediately assess biological response when the patient undergoes surgery [29, 30].

The fourth important trial implicating ER as a predictor of response to endocrine therapy was another window study, by Bundred et al. These authors randomized 90 postmenopausal patients with ER-positive DCIS to one of the following treatments for 2 weeks before surgery: exemestane, an aromatase inhibitor; celecoxib, a COX-2 inhibitor; exemestane and celecoxib; or placebo [30]. Exemestane significantly reduced proliferation compared with placebo; the median reduction was 9%. The effect of exemestane on proliferation was seen regardless of grade, HER2 expression, or PR expression. Celecoxib had no effect on proliferation or apoptosis either when the drug was administered alone or when it was administered in combination with exemestane. Thus, pharmacologic endocrine ablation with exemestane in postmenopausal patients with ER-positive DCIS resulted in a significant reduction in proliferation of cancer cells in the short term. However, the clinical utility of aromatase inhibitors remains to be determined from studies to be reported utilizing this class of drugs in the adjuvant setting [1].

Progesterone Receptor

PR is considered to be as important as ER in *invasive* breast cancer. In *invasive* breast cancer, expression of PR is weakly prognostic with respect to disease-free survival and *also* a predictor of response to endocrine therapy. Among the 28 studies in our review that examined PR expression rate in DCIS, the mean PR expression rate was 59.6% (range: 40–83.3%, Table 2). As with ER, an inverse relationship was observed between PR expression and nuclear grade. Patients with high-grade DCIS were less likely than patients with non-high-grade DCIS to have PR-positive disease [8-10, 12, 14, 15]. One study revealed that PR expression was more common in cribriform DCIS and noncomedo DCIS than in other subtypes [7]. Another study revealed that PR expression was more common in papillary, solid, cribriform, micropapillary, and comedocarcinoma subtypes than in other subtypes [9].

A majority of the studies included in our review investigated the relationship between PR and other biological markers. A direct positive relationship was

observed between PR expression and ER expression [7, 9, 21]. Also, a positive relationship was observed between PR expression and Bcl-2 expression [7, 16, 22]. One study revealed a significant correlation between PR status and p27 status [31]. Several studies reported an inverse relationship between PR expression and HER2 expression [7, 9, 12, 16, 17, 21]. A few studies revealed an inverse relationship between PR expression and p53 expression [7, 16]. In addition, an inverse relationship was observed between PR expression and Ki-67 expression [15, 16].

Thirteen studies (2,051 total patients) in our review evaluated the relationship between PR expression and risk of recurrence, and only two of these studies revealed a significant correlation. In a nested case-control study, Provenzano et al found that PR negativity was independently associated with risk of local-regional recurrence. These authors compared tissues from patients who subsequently developed ipsilateral recurrence (cases) with those from patients who did not develop a recurrence (controls). Patients who developed a local-regional recurrence were more likely than controls to have PR-negative disease (63% vs. 34%, Table 9) [22]. As mentioned previously, Ringberg et al found that a cell biological index that included ER and PR negativity, overexpression of HER2, low Bcl-2 expression, accumulation of p53, nondiploidy, and high Ki-67 expression was a strong predictor of recurrence [16].

Androgen Receptor

Breast cancer is known to be a hormonally dependent carcinoma. Many immunohistochemical studies conducted to date have revealed androgen receptor (AR) to be often co-expressed with ER and PR in breast tumors, but only a few studies have examined the role and significance of AR in breast cancer etiology and prognosis [32, 33]. Among the four studies in our review that examined AR expression rate in DCIS, the mean AR expression rate was 65.8% (range: 37–81%, Table 2). Hanley et al found that unlike expression of the other steroid receptors, ER and PR, expression of AR was slightly higher in high-grade DCIS than in non-high-grade DCIS, although the difference was not significant (93% vs. 89%) [8]. Hanley et al studied patterns of co-expression of ER and PR with AR. These authors reported that 87% of non-high-grade DCIS lesions compared with 30% of high-grade DCIS lesions expressed both AR and ER. The co-expression pattern was similar for PR [8]. Yu et al reported similar findings with respect to co-expression of AR with ER ($P < 0.001$) and PR ($P = 0.035$) [33].

There were only two studies that investigated

the relationship between AR expression and risk of recurrence. Provenzano et al found that AR expression was not a predictor of local recurrence of DCIS [22]. Another study that we identified during our initial search did not meet our selection criteria because too few patients were studied; only 34 patients with DCIS were investigated [33].

Proliferation marker—Ki-67

Information on expression of Ki-67 in DCIS is presented in Table 3. The nuclear antigen Ki-67 is commonly used to assess the proliferation rate of breast cancer tumors, which is a key element of progression of the disease [34]. Our review included 16 studies that reported Ki-67 expression rates in DCIS (Table 3). Barnes et al. reported a median expression of Ki-67 in DCIS as 10.9 - 15.5% (Table 3) [35]. Kuerer et al reported that the mean (\pm standard deviation) percentage of cells staining for Ki-67 was $44.29 \pm 3.42\%$ (Table 3). This expression rate is likely high because all the patients in this series had HER2-positive DCIS, and were more likely to have high-grade lesions [18]. Bundred et al reported that the median percentage of cells staining for Ki-67 at baseline was 13.5–20.7% in the various subgroups in a randomized trial of aromatase-inhibitor therapy in patients with ER-positive DCIS (Table 3) [30].

A few studies revealed high proliferative activity to be comedo DCIS than in DCIS with other architectural patterns [7, 16]. In addition, high proliferative activity was associated with high-grade versus non-high-grade DCIS lesions [12, 16, 36].

Three of the nine studies (1,365 total patients) in our review that evaluated the relationship between Ki-67 expression and the risk of local recurrence concluded that Ki-67 was not significantly associated with disease recurrence (Table 9). The remaining studies showed that Ki-67 was a predictor of local recurrence, either independently or in combination with other factors. Barnes et al, by means of multivariate analysis, reported that Ki-67 was an independent predictor of recurrence (Odds ratio: 1.03; 95% CI: 1.00-1.06; $P=0.038$, Table 9). Patients in this study who had had a recurrence had significantly higher median proliferative activity than patients who had not had a recurrence (15.5% vs. 10.9%, $P=0.005$) [35]. In a similar study conducted by Wilson et al, by means of univariate analysis, patients with recurrence were more likely to exhibit high proliferative activity than patients without recurrence (71.4 % vs. 42.2%, $P=0.006$). However, in the multivariate analysis, Ki-67 was not found to be a predictor of local recurrence. [37]. Kerlikowske et al found that high Ki-67 expression (more than 10% of tumor cells stained) was indi-

vidually associated with recurrence of DCIS in the univariate analysis, but, Ki-67 was not individually associated with subsequent *invasive* tumor. However, the phenotype Ki-67+p16+ and the phenotype Ki-67+p16+COX-2+ were associated with subsequent *invasive* tumor. Ki-67 was individually associated with DCIS recurrence, and the phenotypes Ki-67+ER-, Ki-67+p16+, and Ki-67+p16+COX-2+ were also associated with DCIS recurrence. In the multivariate analysis, the Ki-67+p16+COX-2+ phenotype was a strong predictor of subsequent *invasive* recurrence (HR: 2.2; 95% CI: 1.1-4.5, Table 9). Ki-67+p16+COX-2- (HR: 3.7; 95% CI: 1.7-7.9, Table 9) and ER-HER2+Ki-67+ (HR: 5.8; 95% CI: 2.4-14, Table 9) were two phenotypes that were strong predictors of subsequent DCIS recurrence [24].

Cell Cycle Regulation and Apoptotic Markers

Cyclin D1

Cyclin D1 is one of the many proteins involved in cell cycle regulation, specifically regulation of the mid-G1 phase. It is also responsible for regulating cyclin-dependent kinases [38]. Overexpression of cyclin D1 is commonly seen in breast cancer, but it has also been identified in many other tumors [39, 40]. Among the six studies in our review that examined cyclin D1 expression rate in DCIS, the mean cyclin D1 expression rate was 55.9% (range: 37–70.6%, Table 4).

Oh et al reported a significant correlation between cyclin D1 expression and ER expression [41]. However, Lebeau et al or Millar et al did not find an association between overexpression of cyclin D1 and expression of ER and PR [12, 31].

In our comprehensive search, we identified five studies (443 total patients) that investigated the prognostic significance of cyclin D1. Jirström et al found cyclin D1 to be strongly and inversely related with ipsilateral local recurrence. Patients with low cyclin D1 fraction had a higher risk of ipsilateral local recurrence than patients with high cyclin D1 fraction (32.3% vs. 18.2%, $P=0.002$, Table 9) [42]. Findings from the other four studies indicated no significant correlation between cyclin D1 and risk of local-regional recurrence [21, 31, 38, 43].

Cyclin A

Cyclin A, like cyclin D1, is one of the proteins in the cell cycle responsible for regulating cyclin-dependent kinases [38]. We found only two studies that investigated the role and significance of cyclin A in DCIS. Millar et al reported that 35% of the DCIS lesions in their study expressed cyclin A (Table 4), and Chasle et al did not report an expression rate in their study. These authors found no association

between cyclin A expression and expression of ER or PR or with any histologic features. However, Chasle et al found cyclin A expression to be significantly higher in comedo DCIS than in noncomedo DCIS. In the same study, the investigators found cyclin A expression to be positively correlated with expression of two other biological markers, Ki-67 and p21 [38]. Overall, the investigators did not observe cyclin A to be an independent predictor of local-regional recurrence; however, when they studied global proliferation factor, a combination of Ki-67 and cyclin A, they found this factor to be correlated with local recurrence [38].

Cyclin E

Like cyclin D1 and cyclin A, cyclin E interacts with cyclin-dependent kinases in the late-G1 phase of the cell cycle [44]. We identified one study that investigated the relationship between cyclin E expression and recurrence in DCIS. Jirström et al reported that approximately 25% of the 92 cases of DCIS in their study had high cyclin E expression [42] (Table 9). In the same study, the investigators found that cyclin E was not an independent predictor of disease recurrence [42].

P16

P16 is a tumor suppressor protein that regulates cellular proliferation and growth by acting as a cyclin-dependent kinase 4 inhibitor. In the three studies we identified that examined p16 expression in DCIS, the mean p16 expression rate in DCIS was 33.7% (range: 28–39.3%, Table 4). Gauthier et al did not observe associations between p16 expression and any histologic factors or hormone receptors [45], and the other two studies did not look for associations between p16 expression and other factors.

We identified three studies (576 total patients) that investigated the effect of p16 expression on local recurrence of DCIS. Jirström et al found that p16 was not an independent risk factor for recurrence [42]. Interestingly, Gauthier et al reported that among patients with high p16 expression and high COX-2 expression, patients with high proliferative activity were more likely to have a local-regional recurrence than patients with low proliferative activity [45]. In a follow-up study by the same investigators, Kerlikowske et al, by means of univariate analysis, found p16 to be the only individual marker associated with subsequent *invasive* recurrence (57% vs. 30%, HR: 2.3, Table 9). p16 was also associated with subsequent *invasive* recurrence when it was combined with two other biological markers, Ki-67 expression and COX-2 expression. p16 was also associated with subsequent

DCIS recurrence when it was combined with Ki-67 expression and lack of COX-2 expression. In the multivariate analysis, the p16+COX-2+Ki-67+ triple-positive phenotype was a strong predictor of subsequent *invasive* recurrence (HR: 2.2; 95% CI: 1.1-4.5, Table 9), and the p16+COX-2-Ki-67+ phenotype was a predictor of subsequent DCIS recurrence (HR: 3.7; 95% CI: 2.4-14, Table 9) [24].

P21

P21 is an important protein that plays a role in cell cycle arrest. Studies have shown that p21 can either be a downstream target of p53 or can induce cell arrest independently of p53 [12, 46]. Among the six studies we identified that examined p21 expression rate in DCIS, the mean p21 expression rate was 53.1% (range: 29.6–76%, Table 4). Lebeau et al reported p21 to be significantly associated with nuclear grade and Van Nuys prognostic index; however, Oh et al reported p21-positive DCIS to be associated with well-differentiated histologic grade [12, 41]. With respect to clinicopathological factors, Oh et al also found that p21 expression was associated with noncomedo DCIS and tumors without necrosis [41]. When Lebeau et al studied the relationship between p21 and other biological markers, they found no correlation between p21 expression and p53 expression; however, p21 expression was significantly associated with expression of epidermal growth factor receptor and overexpression of HER2. In addition, p21 expression was correlated with cyclin D1 expression, but only in HER2-positive cases [12]. Kulkarni et al also investigated the relationship between p21 and other biological markers, and they found that p21 expression was significantly correlated with ER and cyclin D1 positivity [21]. Oh et al found p21 expression to be significantly positively correlated with ER expression and significantly inversely correlated with p53 expression [41]. Cornfield et al found no correlation between p21 and the other biological markers tested in their study (ER, PR, HER2, Ki-67, and Bcl-2) [46].

Four studies (365 total patients) in our comprehensive search examined the effect of p21 expression on local recurrence in DCIS. Only one study identified p21 as an independent predictor of recurrence. Provenzano et al reported that patients with local-regional recurrence were more likely than those without recurrence to have p21-positive disease (54% vs. 15%, Odds ratio: 6.0; $P=0.01$, Table 9) [22].

P27

P27 is a cyclin-dependent kinase inhibitor, and it also plays an important role in growth signaling. In recent breast cancer studies, loss of p27 has been as-

sociated with poor patient outcome [31, 47]. Among the three studies in our review that examined p27 expression rate in DCIS, the mean p27 expression rate was 56.8% (range: 46.9–68.5%, Table 4). Jirstrom et al studied the associations between p27 and other biological markers, and they found p27 expression to be significantly correlated with p16 and cyclin D1 expression [42]. Millar et al and Oh et al observed p27 status to be significantly associated with cyclin D1 and ER status [31, 41]. Millar et al also found a significant association between p27 status and PR status [31]. In the same study, no association was observed between p27 positivity and cyclin A positivity or between cyclin A positivity and cyclin D1 positivity. Millar et al and Oh et al found no correlation between p27 and clinicopathological factors [31, 41]. In our comprehensive search, we found only two studies (237 total patients) that investigated the relationship between p27 expression and risk of local recurrence in DCIS. Both studies showed that p27 was not a predictor of local recurrence (Table 9).

P53

P53, a tumor suppressor gene, is responsible for apoptosis in the cell cycle. A mutation of p53 can compromise its function. P53 mutations are common in many cancers and occur in approximately 20% of breast carcinomas [43]. Among the 17 studies in our review that examined p53 expression rate in DCIS, the mean p53 expression rate was 41.4% (range: 11.3–88%, Table 4). Two studies revealed p53 expression to be more likely in DCIS of comedo type [7, 48]. Other studies showed p53 expression to be correlated with high tumor grade [36, 48]. Hieken et al also found that p53 was significantly associated with tumor grade and the presence of necrosis [48].

Several studies investigated the relationship between p53 and other biological markers. Ringberg et al reported p53 expression to be positively correlated with expression of ER, PR, HER2, and Bcl-2; however, Lebeau et al did not find p53 to be associated with either ER or PR [12, 16]. Lebeau et al also noted a correlation between p53 expression and lack of expression of cyclin D1 [12].

We found ten studies (1,355 total patients) that looked at the relationship between p53 expression and local recurrence; only three of these studies showed an association. Ringberg et al used principal components analysis to evaluate a cell biological index that included ER and PR negativity, overexpression of HER2, low Bcl-2 expression, accumulation of p53, nondiploidy, and high Ki-67 expression. P53 when combined with these biological markers was a predictor of local-regional recurrence [16]. The specific

rates of local recurrence associated with particular marker phenotypes were not discernible from the published report; however, the overall rate of recurrence was 25.6% among 121 patients treated with breast-conserving surgery without radiotherapy. Hieken et al reported that DCIS that recurred was more likely than DCIS that did not recur to exhibit strong p53 expression (63% vs. 24%, Table 9) [48]. Similarly, de Roos et al reported p53 expression to be an independent predictor of local recurrence (HR: 3.0, 95% CI: 1.1-8.2, $P=0.036$, Table 9) [43].

Bcl-2

Bcl-2 is one of the apoptosis regulator proteins. A deregulation in apoptosis due to imbalances in Bcl-2 expression has been associated with the pathogenesis of breast cancer [49]. Bcl-2 has been shown to be an independent prognostic biomarker in early-stage breast cancer and nasopharyngeal carcinoma [50, 51]. Among the six studies in our review that examined Bcl-2 expression rate in DCIS, the mean expression rate of Bcl-2 in DCIS among six reported studies was 55.4% (range: 48–58.8%, Table 4). Warnberg et al found a higher expression of Bcl-2 in well-differentiated lesions than in poorly differentiated lesions [36]. Several investigators studied the relationship between Bcl-2 and other biological markers. Ringberg et al and Provenzano et al observed a positive association between Bcl-2 expression and expression of ER and PR [16, 22]. Jirstrom et al also observed a significant negative association between Bcl-2 expression and overexpression of HER2 [42].

We identified three studies (433 total patients) that investigated the association between Bcl-2 expression and risk of local-regional recurrence in DCIS. Cornfield et al found that Bcl-2 expression was not an independent predictor of local recurrence [46]. However, Provenzano et al found that DCIS that recurred was more likely than DCIS that did not recur to be Bcl-2 negative (66% vs. 26%, respectively, Table 9) [22].

Bax

Bax is another protein that plays a role in promoting apoptosis [52]. In gastric carcinoma, colorectal, and pancreatic cancer, negative Bax expression has been shown to be a prognostic predictor [53]. According to Okumura et al, the rate of expression of Bax in DCIS was 71.2% (Table 4) [52]. Bax is one of the less studied proteins in DCIS; we were unable to identify studies that investigated the significance of Bax expression in terms of local recurrence of DCIS.

Survivin

Survivin is another important protein in the cell cycle, responsible for cell cycle regulation and inhibition of apoptosis [52]. Okumura reported an expression rate of survivin in DCIS of 55.8% (Table 4) [52]. Barnes et al reported the expression of survivin according to different staining: nuclear staining, 10%; cytoplasmic staining, 29%; and cytoplasmic and nuclear staining, 29% [54]. Barnes et al reported no significant correlations between survivin and clinicopathological factors. We identified one study that investigated the recurrence risk associated with survivin [54]. In this study, patients with recurrence were more likely than those without recurrence to have co-expression of COX-2 and cytoplasmic survivin (70% vs. 41%, Table 9) [54].

C-myc

C-myc is a proto-oncogene that is known to play a role in proliferation, malignant transformation, and apoptosis. Although c-myc has been associated with poor prognosis in invasive breast cancer, few studies have evaluated c-myc expression levels in DCIS [55]. Altintas et al reported that the expression rate of c-myc in DCIS was 60% (Table 4) [15]. In the same study, Altintas et al found that c-myc was not a predictor of local recurrence of DCIS (Table 9).

Retinoblastoma

Retinoblastoma (Rb) is a tumor suppressor protein that plays an important role in regulating cell growth. Okumura et al reported that the expression rate of Rb in DCIS was 68.6%. In the same study, the investigators found no significant difference in rates of Rb expression between pure DCIS and DCIS with microinvasion [52]. Few studies have evaluated Rb expression levels in DCIS, and no studies investigating the relationship between Rb expression rate and local recurrence of DCIS met our selection criteria.

Angiogenesis-Related Proteins

Vascular endothelial growth factor

Vascular endothelial growth factor (VEGF) is one of the most potent angiogenic factors. It acts as a mitogen for endothelial cells and also increases vascular permeability [48]. Our review included two studies that investigated the expression rate of VEGF in DCIS. The mean value was 89.9% (rates in the two studies were 86.0% and 93.8%, Table 5). No studies in our comprehensive search found VEGF to be correlated with clinicopathological parameters. One study investigated the relationship between VEGF and other biological markers. The investigators found VEGF expression to be significantly associated with COX-2

expression [20]. Only one study, by Hieken et al, investigated the relationship between VEGF expression and local recurrence of DCIS, and the investigators found that VEGF was not an independent predictor of recurrence. In that study, DCIS that recurred was more likely than DCIS that did not recur to express VEGF, but this difference did not reach statistical significance (Table 9) [48].

Heparanase-1

Heparanase-1 (HPR-1), an endo- β -D-glucuronidase, is responsible for cleaving the glycosidic side chains of heparin sulfate proteoglycans. HPR-1 has not been extensively investigated in DCIS, even though an association has been found between HPR-1 and invasive cancer [56]. Maxhimer et al reported that the expression rate of HPR-1 in DCIS was 33.3%. The expression rate of HPR-1 was significantly higher in comedo DCIS than in noncomedo DCIS (75% vs. 14%). The investigators found that tissues from normal breast did not express HPR-1 and that the rate of expression of HPR-1 in DCIS with microinvasion was 83% [56]. In our comprehensive review, we did not identify any studies on the relationship between expression of HPR-1 and recurrence of DCIS.

Epidermal Growth Factor Receptor Family

HER2

Besides the steroid receptors, ER and PR, HER2 is one of the most extensively studied biological markers in DCIS. Studies have found HER2 to be of prognostic significance in invasive cancer; however, its importance in DCIS has yet to be elucidated [57]. Among the 36 studies in our review that examined HER2 expression rate in DCIS, the mean expression rate was 40.1% (range: 9–67%, Table 6). HER2 testing in majority of the studies was different. Some studies assessed HER2 expression based on membrane staining, whereas, other more recent studies assessed HER2 expression from the results of FISH amplification. This complete information is included in Table 6. *Expression rates of human epidermal growth factor receptor (HER) family in DCIS in "definition of positivity."* Most of the studies in our review that investigated HER2 expression rates by subtype found that HER2 expression to be more common in the comedo subtype than in other subtypes of DCIS [7, 9, 17].

Several studies investigated the relationship between HER2 and other biological markers. A few studies showed HER2 to be inversely correlated with ER and PR expression [9, 12, 17, 57]. One study showed HER2 to be positively associated with p53 expression [16]. A few studies showed HER2 overex-

pression to be negatively associated with Bcl-2 expression [16, 22]. The majority of the studies in our comprehensive review showed HER2 overexpression to be positively and significantly correlated with high nuclear grade [9, 10, 12, 17, 18, 22, 36, 57, 58]. However, one study did not show a significant association between HER2 overexpression and grade [20]. One study showed HER2 overexpression to be significantly correlated with proliferative activity [12]. In the same study, HER2 overexpression was significantly associated with p21 status [12].

We identified 15 (2,365 total patients) studies that evaluated the relationship between HER2 expression and local recurrence in DCIS. Eleven of these studies revealed no significant correlation between HER2 and disease recurrence. As mentioned previously, Ringberg et al evaluated a cell biological index that included ER and PR negativity, overexpression of HER2, low Bcl-2 expression, accumulation of p53, nondiploidy, and high Ki-67 expression; this study showed that the index was significantly associated with disease recurrence [16]. Provenzano et al, in a nested case-control study, found HER2 positivity to be individually associated with disease recurrence. In that study, DCIS that subsequently recurred was more likely than DCIS that did not recur to be HER2 positive (41% vs. 12%, Table 9). Kepple et al reported that HER2 positivity was an independent predictor of local-regional recurrence and that HER2 positivity in association with ER positivity was also a predictor of local-regional recurrence. However, it is difficult to interpret the effect of HER2 expression on local recurrence in this study because there were only 37 patients who underwent breast-conserving therapy (with or without radiotherapy), and only four patients had a recurrence [59]. Kerlikowske et al by multivariate analysis did not find HER2 to be an independent predictor of disease recurrence, but they did find in the univariate analysis that HER2 individually and the phenotype HER2+ER-Ki-67+ phenotype were associated with DCIS recurrence. DCIS that recurred was more likely to exhibit the ER-HER2+ phenotype than was DCIS that did not recur (19% vs. 6.4%, respectively, Table 9). In the multivariate analysis, the HER2+ ER-Ki-67+ phenotype was a strong predictor of subsequent DCIS recurrence (OR: 5.8; 95% CI: 2.4-14, Table 9). [24]. Zhou et al evaluated HER2 together with ER and PR and found that basal-like DCIS (negative for ER, PR, and HER2) was more likely to recur than was non-basal-like DCIS (HR: 1.7 vs. 1.8, Table 9). However, this difference did not reach statistical significance [25]. Holmes et al reported HER2 overexpression to be an independent predictor of recurrence. All the patients (n=141) in this study were

treated with lumpectomy alone, and 60 (42.6%) recurrences were reported at a median follow-up of 10.2 years. In the univariate analysis, HER2 overexpression was significantly related to time to recurrence (Table 9). In the multivariate analysis, HER2 positivity (3+) was a strong predictor of recurrence (HR: 1.82, 95% CI: 1.03-3.22, $P=0.041$, Table 9). The overall recurrence rate in this study, 42.6%, is considered to be high and may reflect the fact that the study covered a long time period, with some patients treated as early as 1983 [58].

HER1, HER3, and HER4

Few studies have been conducted to investigate the rates of expression of HER1, HER3, and HER4 in DCIS and the relationship between expression of these markers and risk of local recurrence. The mean HER1 expression rate in DCIS in the three studies included in our review was 23.8% (rates in the individual studies were 13%, 22%, and 36.4%, Table 6). The mean HER3 expression rate in DCIS in the two studies included in our review was 59% (rates in the individual studies were 56% and 62%, Table 6). Finally, the mean HER4 expression rate in DCIS in the two studies that we reviewed was 46% (rates in the individual studies were 37% and 55%, Table 6). No significant correlations were observed between expression of these three biological markers and high-grade vs. low-grade DCIS [15]. Studies also investigated the relationship of HER1, HER2, and HER3 to other markers. One study showed no correlation between these three biological markers and Ki-67 [15]. In the same study, overexpression of c-myc was significantly associated with HER3 and HER4 overexpression; however, no correlation as observed between c-myc overexpression and HER1.

In our comprehensive search, we identified two studies that investigated the recurrence risk associated with the expression of HER1, HER3, and HER4. Altintas et al found that none of these three markers was associated with disease recurrence either individually or in combination with other markers [15]. Barnes et al reported similar findings for HER1 and HER3; however, they observed that HER4 negativity was an independent predictor of recurrence. DCIS that did not recur was more likely than DCIS that did recur to overexpress HER4 (63.3% vs. 35.9%, respectively, Table 9). Both of these studies also investigated patterns of co-expression of HER1-4 with other markers. Barnes et al found that DCIS lesions with the combination of HER2 and HER4 expression were more likely to express ER than were tumors with HER2 expression without HER4 expression (73.2% vs. 52.5%) [35]. Altintas et al found that DCIS lesions

negative for both HER2 and HER4 were more likely to be high-grade lesions in cases (with a recurrence) than in controls (without a recurrence) (90% vs. 57.4%). In the multivariate analysis, expressions of HER1-4 were not independent predictors of local recurrence. In this study, size, margin status, grade/necrosis, and age were the only independent risk factors for local recurrence [15].

Extracellular matrix-related proteins

CD10

CD10 is a zinc-dependent membrane metalloproteinase. This surface biomarker is also known as common acute lymphoblastic leukemia antigen [60]. The mean expression rate of CD10 in DCIS in the two studies included in our review was 22.9% (rates in the individual studies were 18.8% and 27%, Table 7). CD10 expression was found primarily in the myoepithelial cells [60, 61]. In a cohort study, Toussaint et al evaluated normal breast tissue and samples of DCIS and observed expression of CD10 to be lower in DCIS samples than in normal breast tissue [60]. In another cohort study, Witkiewicz et al found a significant association between the expression of CD10 and younger patient age. This was the only study to investigate the relationship between CD10 and other biological markers, and the investigators only found CD10 to be correlated with the glycoprotein secreted protein acidic and rich in cysteine (SPARC) [61]. In our comprehensive search, we identified two studies that investigated the relationship between CD10 expression and risk of recurrence in DCIS. Toussaint et al found low (as opposed to high) CD10 mRNA level in DCIS samples to be significantly associated with risk of recurrence (HR: 2.39; 95% CI: 1.52-3.76, $P=0.001$, Table 9). Of the 154 patients in this study, 20 had had a recurrence (13%) at a median follow-up time of 6 years [60]. Interestingly, the opposite relationship between CD10 and recurrence risk was found by Witkiewicz et al. In their study, strong stromal CD10 expression was significantly associated with disease recurrence (OR: 10.2, 95% CI: 2.7, 37.7, Table 9). This study included 97 patients, and 21 (recurrence rate of 21.6%) had had a recurrence at a median follow-up time of 9.2 years [61].

Secreted Protein Acidic and Rich in Cysteine

SPARC, a 32-kDa glycoprotein, plays an important role in cell matrix interactions and cell cycle progression [61]. We identified only one study that investigated the rate of SPARC expression in DCIS and the relationship between SPARC expression and recurrence in DCIS. Witkiewicz et al reported a SPARC expression rate of 24.7% (Table 7). These in-

vestigators found SPARC to be an independent predictor of recurrence: patients with recurrence were more likely than patients without recurrence to have strong stromal SPARC expression (OR: 3.9, 95% CI: 1.1, 14.3) [61].

Cyclooxygenase-2

COX-2 overexpression has been shown to be upregulated in neoplastic and preneoplastic lesions in the breast [21]. COX-2 is well known to be the rate-limiting enzyme in prostaglandin synthesis. Among the 10 studies in our review that examined COX-2 expression rate in DCIS, the mean COX-2 expression rate was 62.4% (range: 44.4-87.8%, Table 8). Gauthier et al found COX-2 expression to be associated with phospho-p38 staining. P38, a stress-activated kinase, is one of the few markers that has not been investigated in much depth in DCIS [62]. In this cohort of 30 DCIS cases that were available for immunostaining, 61% of the cases overexpressed COX-2; 83% and 87% of the cases, respectively, expressed nuclear and cytoplasmic phospho-p38. All cases in which there was strong COX-2 staining exhibited moderate to high nuclear staining of phospho-p38. However, in cases with high nuclear staining of phospho-p38, the intensity of COX-2 staining varied from low to high [62]. In a few studies, COX-2 was observed to be significantly correlated with high nuclear grade [19, 63, 64]. One study showed COX-2 expression to be correlated with cytoplasmic survivin expression in DCIS [54]. Another study found COX-2 expression to be significantly associated with high Ki-67 expression, ER negativity, and HER2 positivity [19]. However, another study did not find COX-2 overexpression to be significantly associated with HER2 overexpression [21].

We identified four studies that investigated the relationship between COX-2 expression and risk of local-regional recurrence in DCIS. Barnes et al found greater COX-2 expression in DCIS that recurred than in DCIS that did not recur. Multivariate analysis revealed COX-2 expression to be an independent predictor of recurrence in DCIS. In addition, co-expression of COX-2 and cytoplasmic survivin was found in 70% of the recurrent cases, whereas none of the cases lacking expression of both of those biomarkers recurred within 5 years [54]. Kulkarni et al found that COX-2 expression was an independent predictor of recurrence in both the univariate and multivariate analyses. In addition, DCIS that recurred was more likely than DCIS that did not recur to express COX-2 (67% vs. 29%, respectively, Table 9) [21]. Gauthier et al found that high COX-2 expression in combination with high p16 expression and high Ki-67

expression was a risk factor for recurrence [45]. In a follow-up study, Kerlikowske et al did not find COX-2 to be an independent predictor or individually associated with recurrence; however, COX-2 in combination with other phenotypes was a strong predictor and associated with recurrence. In the univariate analysis, cases with subsequent *invasive* recurrence were more likely to express COX-2, p16, and Ki-67 than were cases that did not recur (23% vs. 8.5%, respectively, Table 9) [24]. Cases with DCIS recurrence were more likely than cases without recurrence to have the phenotype COX-2-Ki-67+p16+ (19% vs. 2.6%, Table 9). In the multivariate analysis, the p16, COX-2, and Ki-67 triple-positive phenotype was a strong predictor of subsequent *invasive* recurrence (HR: 2.2; 95% CI: 1.1-4.5; Table 9); whereas the p16+COX-2-Ki-67+ phenotype was a strong predictor of subsequent DCIS recurrence (HR: 3.7; 95% CI: 1.7-7.9; Table 9). [24]. The study by Bundred et al discussed earlier in this article revealed that use of celecoxib, a COX-2 inhibitor, for 2 weeks prior to surgery had no demonstrable effect on proliferation or apoptosis in DCIS [30]. These authors concluded that the use of COX-2 inhibitors would be unlikely to have a therapeutic value in DCIS [30].

Summary

It was difficult to elucidate the prognostic importance of the biomarkers investigated in this comprehensive review because of heterogeneous treatment approaches and often conflicting results. Although the studies in this review provide valuable information on the diagnostic and prognostic significance of the studied markers, another factor that limits our ability to draw conclusions on the basis of the information in this review is the fact that many of the studies reviewed included only small numbers of patients. Other studies included groups of patients treated with different therapies, and in some studies the treatment was inconsistent. In addition, several studies included patients who had received endocrine therapy or radiotherapy, while other studies did not. This heterogeneous treatment makes it hard to assess clinical outcome. In conclusion, novel and key breast cancer biological markers need to be studied prospectively in large cohorts of patient to differentiate indolent from aggressive DCIS and tailor the need and extent of therapies.

Conflict of Interest

The authors have declared that no conflict of interest exists.

References

1. Kuerer HM. Rational individualised selection of adjuvant therapy for ductal carcinoma in situ. *Lancet Oncol* 2010;12:2-3
2. Allegra CJ, Aberle DR, Ganschow P, et al. National Institutes of Health State-of-the-Science Conference statement: Diagnosis and Management of Ductal Carcinoma In Situ September 22-24, 2009. *J Natl Cancer Inst* 2010;102:161-9
3. Madu CO and Lu Y. Novel diagnostic biomarkers for prostate cancer. *J Cancer* 2010;1:150-77
4. Barker PE. Cancer biomarker validation: standards and process: roles for the National Institute of Standards and Technology (NIST). *Ann N Y Acad Sci* 2003;983:142-50
5. Ludwig JA and Weinstein JN. Biomarkers in cancer staging, prognosis and treatment selection. *Nat Rev Cancer* 2005;5:845-56
6. McShane LM, Altman DG, Sauerbrei W, et al. Reporting recommendations for tumor marker prognostic studies (REMARK). *J Natl Cancer Inst* 2005;97:1180-4
7. Albonico G, Querzoli P, Ferretti S, et al. Biological profile of in situ breast cancer investigated by immunohistochemical technique. *Cancer Detect Prev* 1998;22:313-8
8. Hanley K, Wang J, Bourne P, et al. Lack of expression of androgen receptor may play a critical role in transformation from in situ to invasive basal subtype of high-grade ductal carcinoma of the breast. *Hum Pathol* 2008;39:386-92
9. Claus EB, Chu P, Howe CL, et al. Pathobiologic findings in DCIS of the breast: morphologic features, angiogenesis, HER-2/neu and hormone receptors. *Exp Mol Pathol* 2001;70:303-16
10. Bijker N, Peterse JL, Duchateau L, et al. Histological type and marker expression of the primary tumour compared with its local recurrence after breast-conserving therapy for ductal carcinoma in situ. *Br J Cancer* 2001;84:539-44
11. Lebrecht A, Buchmann J, Hefler L, et al. Histological category and expression of hormone receptors in ductal carcinoma in situ of the breast. *Anticancer Res* 2002;22:1909-11
12. Lebeau A, Unholzer A, Amann G, et al. EGFR, HER-2/neu, cyclin D1, p21 and p53 in correlation to cell proliferation and steroid hormone receptor status in ductal carcinoma in situ of the breast. *Breast Cancer Res Treat* 2003;79:187-98
13. Collins LC and Schnitt SJ. HER2 protein overexpression in estrogen receptor-positive ductal carcinoma in situ of the breast: frequency and implications for tamoxifen therapy. *Mod Pathol* 2005;18:615-20
14. Meijnen P, Peterse JL, Antonini N, et al. Immunohistochemical categorisation of ductal carcinoma in situ of the breast. *Br J Cancer* 2008;98:137-42
15. Altintas S, Lambein K, Huizing MT, et al. Prognostic significance of oncogenic markers in ductal carcinoma in situ of the breast: a clinicopathologic study. *Breast J* 2009;15:120-32
16. Ringberg A, Anagnostaki L, Anderson H, et al. Cell biological factors in ductal carcinoma in situ (DCIS) of the breast-relationship to ipsilateral local recurrence and histopathological characteristics. *Eur J Cancer* 2001;37:1514-22
17. DiGiovanna MP, Chu P, Davison TL, et al. Active signaling by HER-2/neu in a subpopulation of HER-2/neu-overexpressing ductal carcinoma in situ: clinicopathological correlates. *Cancer Res* 2002;62:6667-73
18. Kuerer HM, Buzdar AU, Mittendorf EA, et al. Biologic and immunologic effects of preoperative trastuzumab for ductal carcinoma in situ of the breast. *Cancer* 2011;117(1):39-47
19. Boland GP, Butt IS, Prasad R, et al. COX-2 expression is associated with an aggressive phenotype in ductal carcinoma in situ. *Br J Cancer* 2004;90:423-9
20. Perrone G, Santini D, Vincenzi B, et al. COX-2 expression in DCIS: correlation with VEGF, HER-2/neu, prognostic molecu-

- lar markers and clinicopathological features. *Histopathology* 2005;46:561-8
21. Kulkarni S, Patil DB, Diaz LK, et al. COX-2 and PPARgamma expression are potential markers of recurrence risk in mammary duct carcinoma in-situ. *BMC Cancer* 2008;8:36
 22. Provenzano E, Hopper JL, Giles GG, et al. Biological markers that predict clinical recurrence in ductal carcinoma in situ of the breast. *Eur J Cancer* 2003;39:622-30
 23. Roka S, Rudas M, Taucher S, et al. High nuclear grade and negative estrogen receptor are significant risk factors for recurrence in DCIS. *Eur J Surg Oncol* 2004;30:243-7
 24. Kerlikowske K, Molinaro AM, Gauthier ML, et al. Biomarker expression and risk of subsequent tumors after initial ductal carcinoma in situ diagnosis. *J Natl Cancer Inst* 2010;102:627-37
 25. Zhou W, Jirstrom K, Johansson C, et al. Long-term survival of women with basal-like ductal carcinoma in situ of the breast: a population-based cohort study. *BMC Cancer* 2010;10:653
 26. Ma CX, Sanchez CG and Ellis MJ. Predicting endocrine therapy responsiveness in breast cancer. *Oncology (Williston Park)* 2009;23:133-42
 27. Cuzick J, Sestak I, Pinder SE, et al. Effect of tamoxifen and radiotherapy in women with locally excised ductal carcinoma in situ: long-term results from the UK/ANZ DCIS trial. *Lancet Oncol* 2011;12:21-9
 28. Eng-Wong J, Costantino JP and Swain SM. The impact of systemic therapy following ductal carcinoma in situ. *J Natl Cancer Inst Monogr* 2010;2010:200-3
 29. Chen YY, DeVries S, Anderson J, et al. Pathologic and biologic response to preoperative endocrine therapy in patients with ER-positive ductal carcinoma in situ. *BMC Cancer* 2009;9:285
 30. Bundred NJ, Cramer A, Morris J, et al. Cyclooxygenase-2 inhibition does not improve the reduction in ductal carcinoma in situ proliferation with aromatase inhibitor therapy: results of the ERISAC randomized placebo-controlled trial. *Clin Cancer Res* 2010;16:1605-12
 31. Millar EK, Tran K, Marr P, et al. p27KIP-1, cyclin A and cyclin D1 protein expression in ductal carcinoma in situ of the breast: p27KIP-1 correlates with hormone receptor status but not with local recurrence. *Pathol Int* 2007;57:183-9
 32. Birrell SN, Hall RE and Tilley WD. Role of the androgen receptor in human breast cancer. *J Mammary Gland Biol Neoplasia* 1998;3:95-103
 33. Yu Q, Niu Y, Liu N, et al. Expression of androgen receptor in breast cancer and its significance as a prognostic factor. *Ann Oncol* 2010; Epub
 34. de Azambuja E, Cardoso F, de Castro GJr., et al. Ki-67 as prognostic marker in early breast cancer: a meta-analysis of published studies involving 12,155 patients. *Br J Cancer* 2007;96:1504-13
 35. Barnes NL, Khavari S, Boland GP, et al. Absence of HER4 expression predicts recurrence of ductal carcinoma in situ of the breast. *Clin Cancer Res* 2005;11:2163-8
 36. Warnberg F, Nordgren H, Bergkvist L, et al. Tumour markers in breast carcinoma correlate with grade rather than with invasiveness. *Br J Cancer* 2001;85:869-74
 37. Wilson GR, Cramer A, Welman A, et al. Activated c-SRC in ductal carcinoma in situ correlates with high tumour grade, high proliferation and HER2 positivity. *Br J Cancer* 2006;95:1410-4
 38. Chasle J, Delozier T, Denoux Y, et al. Immunohistochemical study of cell cycle regulatory proteins in intraductal breast carcinomas--a preliminary study. *Eur J Cancer* 2003;39:1363-9
 39. Zhou CJ, Zhang QH, Zhang TG, et al. Expression of ER, Ki-67 and cyclinD1 in the pre-cancerous breast of Chinese patients. *Pathol Oncol Res* 2009;15:153-8
 40. Guo LL, Gao P, Wu YG, et al. Alteration of cyclin D1 in Chinese patients with breast carcinoma and its correlation with Ki-67, pRb, and p53. *Arch Med Res* 2007;38:846-52
 41. Oh YL, Choi JS, Song SY, et al. Expression of p21Waf1, p27Kip1 and cyclin D1 proteins in breast ductal carcinoma in situ: Relation with clinicopathologic characteristics and with p53 expression and estrogen receptor status. *Pathol Int* 2001;51:94-9
 42. Jirstrom K, Ringberg A, Ferno M, et al. Tissue microarray analyses of G1/S-regulatory proteins in ductal carcinoma in situ of the breast indicate that low cyclin D1 is associated with local recurrence. *Br J Cancer* 2003;89:1920-6
 43. de Roos MA, de Bock GH, de Vries J, et al. p53 overexpression is a predictor of local recurrence after treatment for both in situ and invasive ductal carcinoma of the breast. *J Surg Res* 2007;140:109-14
 44. Steeg PS and Zhou Q. Cyclins and breast cancer. *Breast Cancer Res Treat* 1998;52:17-28
 45. Gauthier ML, Berman HK, Miller C, et al. Abrogated response to cellular stress identifies DCIS associated with subsequent tumor events and defines basal-like breast tumors. *Cancer Cell* 2007;12:479-91
 46. Cornfield DB, Palazzo JP, Schwartz GF, et al. The prognostic significance of multiple morphologic features and biologic markers in ductal carcinoma in situ of the breast: a study of a large cohort of patients treated with surgery alone. *Cancer* 2004;100:2317-27
 47. Alkarain A, Jordan R and Slingerland J. p27 deregulation in breast cancer: prognostic significance and implications for therapy. *J Mammary Gland Biol Neoplasia* 2004;9:67-80
 48. Hieken TJ, Farolan M, D'Alessandro S, et al. Predicting the biologic behavior of ductal carcinoma in situ: an analysis of molecular markers. *Surgery* 2001;130:593-600
 49. Rehman S, Crow J and Revell PA. Bax protein expression in DCIS of the breast in relation to invasive ductal carcinoma and other molecular markers. *Pathol Oncol Res* 2000;6:256-63
 50. Dawson SJ, Makretsov N, Blows FM, et al. BCL2 in breast cancer: a favourable prognostic marker across molecular subtypes and independent of adjuvant therapy received. *Br J Cancer* 2010;103:668-75
 51. Chen MK, Yang SF, Lai JC, et al. Expression of bcl-2 correlates with poor prognosis and modulates migration of nasopharyngeal carcinoma cells. *Clin Chim Acta* 2010;411:400-5
 52. Okumura Y, Yamamoto Y, Zhang Z, et al. Identification of biomarkers in ductal carcinoma in situ of the breast with microinvasion. *BMC Cancer* 2008;8:287
 53. Anagnostopoulos GK, Stefanou D, Arkoumani E, et al. Expression of Bax protein in gastric carcinomas. A clinicopathological and immunohistochemical study. *Acta Gastroenterol Belg* 2007;70:285-9
 54. Barnes N, Haywood P, Flint P, et al. Survivin expression in in situ and invasive breast cancer relates to COX-2 expression and DCIS recurrence. *Br J Cancer* 2006;94:253-8
 55. Perez EA, Jenkins RB, Dueck AC, et al. C-MYC Alterations and Association With Patient Outcome in Early-Stage HER2-Positive Breast Cancer From the North Central Cancer Treatment Group N9831 Adjuvant Trastuzumab Trial. *J Clin Oncol* 2011; 29(6):651-9
 56. Maxhimer JB, Pesce CE, Stewart RA, et al. Ductal carcinoma in situ of the breast and heparanase-1 expression: a molecular explanation for more aggressive subtypes. *J Am Coll Surg* 2005;200:328-35
 57. Latta EK, Tjan S, Parkes RK, et al. The role of HER2/neu overexpression/amplification in the progression of ductal carcinoma in situ to invasive carcinoma of the breast. *Mod Pathol* 2002;15:1318-25

58. Holmes P, Lloyd J, Chervoneva I, et al. Prognostic markers and long-term outcomes in ductal carcinoma in situ of the breast treated with excision alone. *Cancer* 2011; Epub
59. Kepple J, Henry-Tillman RS, Klimberg VS, et al. The receptor expression pattern in ductal carcinoma in situ predicts recurrence. *Am J Surg* 2006;192:68-71
60. Toussaint J, Durbecq V, Altintas S, et al. Low CD10 mRNA expression identifies high-risk ductal carcinoma in situ (DCIS). *PLoS One* 2010;5: e12100
61. Witkiewicz AK, Freydin B, Chervoneva I, et al. Stromal CD10 and SPARC expression in ductal carcinoma in situ (DCIS) patients predicts disease recurrence. *Cancer Biol Ther* 2010;10:391-6
62. Gauthier ML, Pickering CR, Miller CJ, et al. p38 regulates cyclooxygenase-2 in human mammary epithelial cells and is activated in premalignant tissue. *Cancer Res* 2005;65:1792-9
63. Tan KB, Yong WP and Putti TC. Cyclooxygenase-2 expression: a potential prognostic and predictive marker for high-grade ductal carcinoma in situ of the breast. *Histopathology* 2004;44:24-8
64. de la Torre J, Sabadell MD, Rojo F, et al. Cyclo-oxygenase type 2 is dysregulated in breast ductal carcinoma in situ and correlates with poor outcome. *Eur J Obstet Gynecol Reprod Biol* 2010;151:72-6
65. Bryan BB, Schnitt SJ and Collins LC. Ductal carcinoma in situ with basal-like phenotype: a possible precursor to invasive basal-like breast cancer. *Mod Pathol* 2006;19:617-21
66. Livasy CA, Perou CM, Karaca G, et al. Identification of a basal-like subtype of breast ductal carcinoma in situ. *Hum Pathol* 2007;38:197-204
67. Tamimi RM, Baer HJ, Marotti J, et al. Comparison of molecular phenotypes of ductal carcinoma in situ and invasive breast cancer. *Breast Cancer Res* 2008;10:R67
68. Roses RE, Paulson EC, Sharma A, et al. HER-2/neu overexpression as a predictor for the transition from in situ to invasive breast cancer. *Cancer Epidemiol Biomarkers Prev* 2009;18:1386-9
69. Suzuki J, Chen YY, Scott GK, et al. Protein acetylation and histone deacetylase expression associated with malignant breast cancer progression. *Clin Cancer Res* 2009;15:3163-71
70. Yu KD, Wu LM, Liu GY, et al. Different Distribution of Breast Cancer Subtypes in Breast Ductal Carcinoma in situ (DCIS), DCIS with Microinvasion, and DCIS with Invasion Component. *Ann Surg Oncol* 2010; 18(5):1342-8
71. Behling KC, Tang A, Freydin B, et al. Increased SIAH expression predicts ductal carcinoma in situ (DCIS) progression to invasive carcinoma. *Breast Cancer Res Treat* 2010; Epub
72. Menter DG, Hoque A, Motiwala N, et al. Computerized image analysis of Ki-67 in ductal breast carcinoma in situ. *Anal Quant Cytol Histol* 2001;23:218-28
73. Hoque A, Sneige N, Sahin AA, et al. Her-2/neu gene amplification in ductal carcinoma in situ of the breast. *Cancer Epidemiol Biomarkers Prev* 2002;11:587-90
74. Stackiewicz R, Paran H, Bernheim J, et al. Prognostic significance of HER-2/neu expression in patients with ductal carcinoma in situ. *Isr Med Assoc J* 2010;12:290-5

Tables

Table 1. Classification scheme of biological markers evaluated in the comprehensive review

Steroid receptors	Estrogen receptor, progesterone receptor, and androgen receptor
Proliferation marker	Ki-67
Cell cycle regulation and apoptotic markers	cyclin D1, cyclin A, cyclin E, p16, p21, p27, p53, Bcl-2, Bax, Survivin, c-myc, and retinoblastoma
Angiogenesis related proteins	Vascular endothelial growth factor and heparanase-1
Epidermal growth factor receptor family	HER1, HER2, HER3, and HER4
Extracellular matrix related proteins	CD10 Secreted protein acidic and rich in cysteine
Other biological marker	COX-2

Table 2. Expression rates of steroid receptors in DCIS

Biomarker	First author and reference	Year	No. of samples	Expression rate, %	Definition of positivity
<i>Estrogen receptor (ER)</i>					
	Albonico [7]	1998	62	28.8-92.6	Nuclear staining in at least 10% of tumor cells. 28.8 (comedo DCIS) 78.9 (noncomedo DCIS) 92.6 (cribriform DCIS)
	Claus [9]	2001	219	60	No cut-off value was mentioned in the paper.
	Bijker [10]	2001	116	62.9	Any ER expression.
	Ringberg [16]	2001	187	60	Nuclear staining in more than 10% of tumor cells.
	Wärnberg [36]	2001	194	68	Nuclear staining in at least 10% of tumor cells.
	Oh [41]	2001	49	65.3	Nuclear staining in more than 5% of tumor cells.
	Lebrecht [11]	2002	120	71.7	Moderate or strong nuclear staining in tumor cells.

DiGiovanna [17]	2002	219	60	Nuclear staining in more than 10% of tumor cells.
Provenzano [22]	2003	95	49	Nuclear staining in at least 10% of tumor cells.
Lebeau [12]	2003	45	55	Nuclear staining in at least 10% of tumor cells.
Roka [23]	2004	190	57.9	Nuclear staining in at least 10% of tumor cells.
Boland [19]	2004	187	59.7	Nuclear staining in at least 5% of tumor cells.
Barnes [35]	2005	129	66.4	Nuclear staining in at least 5% of tumor cells.
Perrone [20]	2005	49	75.5	Nuclear staining in more than 10% of tumor cells.
Collins [13]	2005	148	77	Nuclear staining in at least 10% of tumor cells.
Gauthier [62]	2005	30	78.9	No cut-off value was mentioned in the paper.
Bryan [65]	2006	66	56	Nuclear staining in at least 10% of tumor cells. All cases were high-nuclear-grade DCIS.
Kepple [59]	2006	94	74	No cut-off value was mentioned in the paper
Wilson [37]	2006	129	65.1	Nuclear staining in at least 5% of tumor cells.
Barnes [54]	2006	161	63.6	Nuclear staining in at least 5% of tumor cells.
Millar [31]	2007	60	58.3	Nuclear staining in more than 10% of tumor cells.
Livasy [66]	2007	245	70	Allred score above 2.
Meijnen [14]	2008	163	68	Any ER expression.
Hanley [8]	2008	90	30 (high-grade DCIS); 96 (non-high-grade DCIS)	Nuclear staining in at least 10% of tumor cells.
Tamimi [67]	2008	272	74	Nuclear staining in more than 10% of tumor cells.
Okumura [52]	2008	52	73.1	Nuclear staining in more than 10% of tumor cells.
Kulkarni [21]	2008	69	77.8	Nuclear staining in at least 10% of tumor cells.
Roses [68]	2009	84	76.2	Allred score less than or equal to 3.
Suzuki [69]	2009	58	60	Nuclear staining in at least 1% of tumor cells.
Altintas [15]	2009	159	72	Nuclear staining in at least 10% of tumor cells.
Yu K [70]	2010	271	67.3	Results of immunohistochemical assays were evaluated using a proportion score and an intensity score. Tumors that scored over 1 were scored as positive.
Kuerer [18]	2010	69	81	No cut-off value was mentioned in the paper.
Kerlikowske [24]	2010	329	77.9	Nuclear staining in at least 10% of tumor cells.
Behling [71]	2010	65	78	No cut-off value was mentioned in the paper.
Witkiewicz [61]	2010	97	96.6	Status obtained from the pathology reports.
Holmes [58]	2011	141	80.9	Nuclear staining was assessed in tumors and assigned a score. Tumors with staining in 0-9% of cells were given a score of 3, tumors with staining in 10-79% of cells were given a score of 2, and tumors with staining in 80% or more of cells were given a score of 1. Scores 1 and 2 were considered to be positive.
<i>Progesterone receptor (PR)</i>				
Albonico [7]	1998	62	28.2-89.1	Nuclear staining in at least 10% of tumor cells. 28.2 (comedo DCIS) 75.3 (noncomedo DCIS) 89.1 (cribriform DCIS)
Claus [9]	2001	219	62	No cut-off value was mentioned in the paper.
Bijker [10]	2001	116	47.1	Any PR expression.
Ringberg [16]	2001	187	43	Nuclear staining in more than 10% of tumor cells.
Wärnberg [36]	2001	194	43	Nuclear staining in at least 10% of tumor cells.
DiGiovanna [17]	2002	219	62	Nuclear staining in more than 10% of tumor cells.
Provenzano [22]	2003	95	48	Nuclear staining in at least 10% of tumor cells.
Lebeau [12]	2003	45	50	Nuclear staining in at least 10% of tumor cells.
Roka [23]	2004	190	41	Nuclear staining in at least 10% of tumor cells.
Perrone [20]	2005	49	65.3	Nuclear staining in more than 10% of tumor cells.
Gauthier [62]	2005	30	70.8	No cut-off value was mentioned in the paper.
Bryan [65]	2006	66	40	Nuclear staining in at least 10% of tumor cells. All cases were high-nuclear-grade DCIS.
Kepple [59]	2006	94	56	No cut-off value was mentioned in the paper.

Millar [31]	2007	60	56.1	Nuclear staining in more than 10% of tumor cells.
Meijnen [14]	2008	163	46	Any PR expression.
Hanley [8]	2008	90	23 (high-grade DCIS); 81 (non-high-grade DCIS)	Nuclear staining in at least 10% of tumor cells.
Tamimi [67]	2008	272	59.9	Nuclear staining in more than 10% of tumor cells.
Okumura [52]	2008	52	71.2	Nuclear staining in more than 10% of tumor cells.
Kulkarni [21]	2008	69	75.9	Nuclear staining in more than 10% of tumor cells.
Roses [68]	2009	84	68.3	Allred score less than or equal to 3.
Suzuki [69]	2009	58	55	Nuclear staining in at least 1% of tumor cells.
Altintas [15]	2009	159	65	Nuclear staining in at least 10% of tumor cells.
Yu K [70]	2010	271	52.7	Results of immunohistochemical assays were evaluated using a proportion score and an intensity score. Tumors that scored over 1 were scored as positive.
Bundred [30]	2010	90	72	Nuclear staining in more than 5% of tumor cells. All cases were ER-positive DCIS.
Kerlikowske [24]	2010	329	74.5	Nuclear staining in at least 10% of tumor cells.
Behling [71]	2010	65	71.2	No cut-off value was mentioned in the paper.
Witkiewicz [61]	2010	97	83.3	Status obtained from pathology reports.
Holmes [58]	2011	141	70.9	Nuclear staining was assessed in tumors and assigned a score. Tumors with staining in 0-9% of cells were given a score of 3, tumors with staining in 10-79% of cells were given a score of 2, and tumors with staining in 80% or more of cells were given a score of 1. Scores 1 and 2 were considered to be positive.
<i>Androgen receptor (AR)</i>				
Provenzano [22]	2003	95	81	Nuclear staining in at least 10% of tumor cells.
Meijnen [14]	2008	163	37	Strong nuclear staining in more than 10% of tumor cells.
Hanley [8]	2008	90	89 (non-high-grade DCIS); 93 (high-grade DCIS)	Nuclear staining in at least 10% of tumor cells.
Yu Q. [33]	2010	34	79.4	Allred score of 2.

Table 3. Expression rates of proliferation marker Ki-67 in DCIS

Biomarker	First author and reference	Year	No. of samples	Expression rate, %	Comments
<i>Ki-67</i>					
	Albonico [7]	1998	62	3.4-65.4	Tumors with nuclear staining in more than 13% of cells were scored as positive. 65.4 (comedo DCIS) 7.5 (noncomedo DCIS) 3.4 (cribriform DCIS)
	Ringberg [16]	2001	187	42	42% of cases exhibited staining in more than 10% of the tumor cells.
	Menter [72]	2001	200	0.99-2.59	Ki-67 labeling index was determined by computerized image analysis. The range of Ki-67 labeling index in DCIS nuclear grades I-III was 0.99-2.59.
	Wärnberg [36]	2001	194	19	19% of cases exhibited staining in at least 10% of the tumor cells.
	Lebeau [12]	2003	45	45.5	45.5% of cases exhibited staining in more than 10% of the tumor cells.
	Boland [19]	2004	187	49.2	The percentage of positively stained nuclei (at least 1,000 cells were counted for each case) was determined using a grid graticule and cell counter at X400 magnification. 50.8% of cases exhibited staining in at least 10% of tumor cells.
	Barnes [35]	2005	129	10.9 (non-recurrent)	The percentage of positively stained nuclei (at least 1,000 cells were counted for each case) was determined using a

				DCIS); 15.5 (recur- rent DCIS)	grid graticule and cell counter at X400 magnification. The median percentage of cells with Ki-67 expression was 10.9-15.5%
Barnes [54]	2006	161	8.7-14.4		The percentage of positively stained nuclei (at least 1,000 cells were counted for each case) was determined using a grid graticule and cell counter at X400 magnification. The median percentage of cells with Ki-67 expression was 8.7-14.4%.
Wilson [37]	2006	129	50.4		The percentage of positively stained nuclei (at least 1,000 cells were counted for each case) was determined using a grid graticule and cell counter at X400 magnification. 50.4% of cases exhibited staining in at least 10% of tumor cells.
Gauthier [45]	2007	70	37.1		37.1% of cases exhibited staining in more than 10% of the tumor cells.
Livasy [66]	2007	245	36		36% of cases exhibited staining in more than 10% of the tumor cells.
Okumura [52]	2008	52	17.9		The percentage of cancer cells with positively stained nuclei was determined. The mean percentage Ki-67 staining was $17.9 \pm 1.5\%$.
Altintas [15]	2009	159	36		36% of cases exhibited staining in more than 10% of the tumor cells.
Kuerer [18]	2010	69	44.29 ± 3.42		No cut-off value was mentioned in the paper. The mean (\pm standard deviation) percentage of cells with Ki-67 staining was $44.29 \pm 3.42\%$. Only patients with HER2-positive DCIS were included in the study.
Kerlikowske [24]	2010	329	47.5		47.5% of cases exhibited staining in more than 10% of the tumor cells, and thus 47.5% was set as the median value (positive cells divided by the number of positive plus negative cells) to divide cases with low and high proliferative activity.
Bundred [30]	2010	90	13.5-20.7		Ki-67 scores were calculated as the percentage of positively stained nuclei. The median percentages in each of the different treatment subgroups in this randomized trial were reported before patients were given aromatase inhibitor therapy. All patients had ER-positive DCIS.

Table 4. Expression rates of cell cycle regulation and apoptotic markers in DCIS

Biomarker	First author and reference	Year	No. samples	of	Expression rate, %	Definition of positivity
<i>cyclin D1</i>	Oh [41]	2001	49		59.2	Moderate or strong nuclear staining in more than 10% of tumor cells.
	Lebeau [12]	2003	45		48.8	Nuclear staining in more than 10% of tumor cells.
	Chasle [38]	2003	50		37	The percentage of marked nuclei was determined for 300-400 nuclei in the most positive foci. The mean for cyclin D1 was reported to be 37%.
	Millar [31]	2007	60		58	Nuclear staining in more than 5% of tumor cells.
	Okumura [52]	2008	52		70.6	Nuclear staining in more than 10% of tumor cells.
	Kulkarni [21]	2008	69		61.5	Nuclear staining in at least 10% of tumor cells.
<i>cyclin A</i>	Millar [31]	2007	60		35	Nuclear staining in more than 10% of tumor cells.
<i>cyclin E</i>	Jirström [42]	2003	92		25	A mean value was used as a cut-off to divide expression of cyclin E.
<i>p16</i>	Jirström [42]	2003	92		37.5	A mean value was used as a cut-off to divide expression of p16.
	Gauthier [45]	2007	70		28	Allred score of at least 2.
	Kerlikowske [24]	2010	329		39.3	On a 3-point scale, a score of at least 2.
<i>p21</i>	Oh [41]	2001	49		67.3	Moderate or strong nuclear staining in more than 10% of tumor cells.
	Provenzano [22]	2003	95		34	On a 6-point scale, a score of 4-6 (moderate or strong stain-

	Lebeau [12]	2003	45	42.2	ing).
	Chasle [38]	2003	50	76	Nuclear staining in more than 10% of tumor cells.
	Okumura [52]	2008	52	69.2	The percentage of marked nuclei was determined for 300-400 nuclei in the most positive foci.
	Kulkarni [21]	2008	69	29.6	Nuclear staining in more than 10% of tumor cells.
<i>p27</i>	Oh [41]	2001	49	46.9	Nuclear staining in at least 10% of tumor cells.
	Jirstrom [42]	2003	92	68.5	Moderate or strong nuclear staining in more than 10% of tumor cells.
	Millar [31]	2007	60	55	Nuclear and cytoplasmic staining intensity were evaluated using a 3-point semiquantitative scoring scale (0=none, 1=weak, 2=moderate, 3=strong).
<i>p53</i>	Albonico [7]	1998	62	7.0-37.3	Nuclear staining in more than 50% of tumor cells.
	Bijker [10]	2001	116	20	Nuclear staining in more than 5% of tumor cells. 37.3 (comedo DCIS) 17.2 (noncomedo DCIS) 7.0 (cribriform DCIS)
	Ringberg [16]	2001	187	26	On an 8-point scale, a score of at least 5.
	Hieken [48]	2001	103	27	Nuclear staining in more than 10% of tumor cells.
	Wärnberg [36]	2001	194	40	Strong nuclear staining in more than 10% of tumor cells.
	Oh [41]	2001	49	30.6	Any p53 expression.
	Provenzano [22]	2003	95	60	Nuclear staining in more than 5% of tumor cells.
	Lebeau [12]	2003	45	25	On a 6-point scale, a score of 4-6 (moderate or strong staining).
	Chasle [38]	2003	50	88	Moderate or strong nuclear staining in more than 40% of tumor cells.
	Roka [23]	2004	190	57.2	The percentage of marked nuclei was determined for 300-400 nuclei in the most positive foci.
	Perrone [20]	2005	49	26.5	Nuclear staining in at least 10% of tumor cells.
	Kepple [59]	2006	94	50	Nuclear staining in more than 10% of tumor cells.
	Livasy [66]	2007	245	31	No cut-off value was mentioned in the paper.
	Meijnen [14]	2008	163	26	Nuclear or nuclear and cytoplasmic staining in more than 10% of tumor cells.
	Okumura [52]	2008	52	76.5	Nuclear staining in more than 25% of tumor cells.
	Kulkarni [21]	2008	69	67.9	Nuclear staining in more than 10% of tumor cells.
	Kerlikowske [24]	2010	329	11.3	Nuclear staining in at least 10% of tumor cells.
<i>Bcl-2</i>	Albonico [7]	1998	62	35.7-100	Nuclear staining in more than 10% of tumor cells. 35.7 (comedo DCIS) 100 (noncomedo DCIS) 100 (cribriform DCIS)
	Ringberg [16]	2001	187	56	Cytoplasm staining in more than 10% of tumor cells.
	Wärnberg [36]	2001	194	48	Nuclear staining in at least 10% of tumor cells.
	Provenzano [22]	2003	95	50	On a 6-point scale, a score of 4-6 (moderate or strong staining).
	Meijnen [14]	2008	163	64	Weak cytoplasmic staining in more than 10% of tumor cells.
	Okumura [52]	2008	52	58.8	Nuclear staining in more than 5% of tumor cells.
<i>Bax</i>	Okumura [52]	2008	52	71.2	Nuclear staining in more than 5% of tumor cells.
<i>Survivin</i>	Barnes [54]	2006	161	10 (nuclear staining alone); 29% (cytoplasmic staining alone); 29% (cytoplasmic and nuclear staining)	Survivin staining was scored for both cytoplasmic and nuclear staining. For cytoplasmic staining, a score of at least 2 was considered positive; for nuclear staining, the proportion of positive cells out of at least 1,000 was determined.

<i>C-myc</i>	Okumura [52]	2008	52	55.8	Nuclear staining in more than 5% of tumor cells.
	Altintas [15]	2009	159	60	Score of at least 2.
<i>Rb</i>	Okumura [52]	2008	52	68.6	Nuclear staining in more than 10% of tumor cells.

Table 5. Expression rates of angiogenesis related proteins in DCIS

Biomarker	First author and reference	Year	No. of samples	Expression rate, %	Definition of positivity
<i>Vascular endothelial growth factor (VEGF)</i>					
	Hieken [48]	2001	103	86	Cytoplasmic and/or membrane staining in more than 10% of tumor cells.
	Perrone [20]	2005	49	93.8	Staining (defined as appropriate brown staining in the tumor cell cytoplasm) in more than 10% of tumor cells.
<i>Heparanase-1 (HPR1)</i>					
	Maxhimer [56]	2005	45	42.2	Nuclear staining in more than 20% of tumor cells.

Table 6. Expression rates of human epidermal growth factor receptor (HER) family in DCIS

Biomarker	First author and reference	Year	No. of samples	Expression rate, %	Definition of positivity
<i>Human epidermal growth factor receptor 1 (HER1)</i>					
	Lebeau [12]	2003	45	36.4	Any distinctive membrane staining of intraductal tumor cells.
	Bryan [65]	2006	66	22	Any cytoplasmic and/or membranous staining of tumor cells.
	Altintas [15]	2009	159	13	Score of at least 2
<i>HER2/neu (HER2)</i>					
	Albonico [7]	1998	62	1.0–72.8	Staining in more than 10% of tumor cells. 72.8 (comedo DCIS) 10.8 (noncomedo DCIS) 1.0 (cribriform DCIS)
	Claus [9]	2001	219	28	On a 4-point scale, any score other than 0.
	Bijker [10]	2001	116	46.2	Any expression.
	Ringberg [16]	2001	187	54	Membrane staining in more than 10% of tumor cells.
	Wärnberg [36]	2001	194	55	Moderate or strong membrane staining in at least 30% of tumor cells or complete membrane staining in more than 60% of tumor cells regardless of the intensity of the staining.
	Latta [57]	2002	91	34.1	On an 8-point scale, an IHC score of at least 5. In addition, a HER2/CEP17 ratio of at least 2 was considered positive for HER2/neu gene amplification.
	DiGiovanna [17]	2002	219	28	On a 4-point scale, any score other than 0.
	Hoque [73]	2002	100	40	HER2 gene amplification was analyzed by FISH. A ratio of greater than 2.0 was considered indicative of HER2 gene amplification.
	Provenzano [22]	2003	95	32	Strong staining (equivalent to a score of 3+ with the DakoCytomation HercepTest).
	Lebeau [12]	2003	45	46.7	On a 3-point scale, a score of greater than 2. Scoring was based on the positive staining of the cell membrane.
	Roka [23]	2004	190	41.3	Nuclear staining in more than 30% of tumor cells.
	Boland [19]	2004	187	54.5	On a 3-point scale, a score of at least 2.
	Barnes [35]	2005	129	65	Score of at least 2.
	Perrone [20]	2005	49	66.7	Complete membrane staining in more than 10% of tumor cells.
	Collins [13]	2005	148	28	Strong membrane staining in more than 10% of tumor cells (equivalent to a score of 3+ with the DakoCytomation HercepTest).

	Bryan [65]	2006	66	67	Membrane staining in more than 10% of tumor cells (equivalent to a score of 3+ in the DakoCytomation HercepTest). All cases were high-nuclear-grade DCIS.
	Kepple [59]	2006	94	27	No cut-off value was mentioned in the paper.
	Wilson [37]	2006	129	64.8	On a 3-point scale, a score of at least 2.
	Barnes [54]	2006	161	64.7	Score of at least 2.
	Livasy [66]	2007	245	25	3+ intensity with DAB chromogen and 2+ or 3+ intensity with the SG chromogen in more than 10% of tumor cells.
	Meijnen [14]	2008	163	39	Strong membranous staining in more than 10% of tumor cells.
	Hanley [8]	2008	90	9 (non-high-grade DCIS); 55 (high-grade DCIS)	Membrane staining of 3+.
	Tamimi [67]	2008	272	27.2	Moderate or strong membrane staining (2+ or higher on a 3-point scale) in at least 10% of tumor cells.
	Okumura [52]	2008	52	17.3	On a 3-point scale, a score of 3 (strong staining).
	Kulkarni [21]	2008	69	60.4	On a 3-point scale, 3+ staining in more than 10% of tumor cells.
	Roses [68]	2009	84	28.6	Membranous staining of 3+ in any tumor cell or membranous staining of 2+ in more than 10% of tumor cells with fluorescence in situ hybridization evidence of HER2 gene amplification.
	Suzuki [69]	2009	58	9	On a 3-point scale, a score of 3 or positive for HER2 gene amplification when HER2/CEP17 greater than 2.2.
	Altintas [15]	2009	159	40	Score of at least 2.
	Yu K [70]	2010	271	33.7	Results of immunohistochemical assays were evaluated using a proportion score and an intensity score. HER2 status was defined as positive for scores of 9-12.
	Stackiewicz [74]	2010	84	44	Moderate or strong membrane staining (2+ or higher on a 3-point scale) in more than 10% of tumor cells.
	Kuerer [18]	2010	69	35	On a 3-point scale, a 3+ score was considered to be positive by IHC or positive for HER2 gene amplification when HER2/CEP17 ratio greater than 2.0 by FISH.
	Bundred [30]	2010	90	32	On a 3-point scale, a score of greater than 2. All cases were ER-positive DCIS.
	Kerlikowske [24]	2010	329	18.2	Moderate or strong membrane staining (2+ or higher) in at least 10% of tumor cells.
	Behling [71]	2010	65	50	No cut-off value was mentioned in the paper.
	Witkiewicz [61]	2010	97	33.3	Status obtained from pathology reports.
	Holmes [58]	2011	141	27.7	On a 3-point scale, a score of 3.
<i>HER3</i>					
	Barnes [35]	2005	129	56	Score of at least 2.
	Altintas [15]	2009	159	62	Score of at least 2.
<i>HER4</i>					
	Barnes [35]	2005	129	55	Score of at least 2.
	Altintas [15]	2009	159	37	Score of at least 2.

FISH, fluorescence in situ hybridization; IHC, immunohistochemical.

Table 7. Expression rates of extracellular matrix-related proteins in DCIS

Biomarker	First author and reference	Year	No. of samples	Expression rate, %	Definition of positivity
<i>CD10</i>					
	Toussaint [60]	2010	154	27	CD10 scoring was based on expression and intensity values. A high score was determined to be equivalent to 6.
	Witkiewicz [61]	2010	97	18.8	On a 3-point scale, a score of 2 (defined as strong staining of at least 30% of stromal cells).
<i>SPARC</i>					
	Witkiewicz [61]	2010	97	24.7	On a 3-point scale, a score of 2 (defined as strong staining of at least 30% of stromal cells).

Table 8. Expression rate of COX-2 in DCIS

Biomarker	First author and reference	Year	No. of samples	Expression rate, %	Definition of positivity
	Tan [63]	2004	51	80	On a 9-point scale, 0 was considered to be nil; 1-3 was considered to be mild; 4-6 was considered to moderate; 7-9 was considered to be strong. Only unequivocal cytoplasmic staining was regarded as positive.
	Boland [19]	2004	187	67	On a 3-point scale, a score of at least 2. Expression was based on the extent and intensity of epithelial cell staining.
	Perrone [20]	2005	49	87.8	On a 12-point scale for immunoreactivity, an immunohistochemistry score of 9-12 was considered strong, 5-8 was considered moderate, 1-4 was considered weak, and 0 was considered negative.
	Gauthier [62]	2005	30	60.7	No cut-off value was mentioned in the paper.
	Barnes [54]	2006	161	72	Score of at least 2.
	Gauthier [45]	2007	70	55.7	Allred score of at least 2.
	Kulkarni [21]	2008	69	45.8	Cytoplasmic granular staining in more than 10% of tumor cells.
	Bundred [30]	2010	90	58	A minimum of 500 cells was investigated across randomly selected areas of DCIS at a magnification of x 400 using a grid graticule and cell counter for each of the two sections. All cases were ER-positive DCIS.
	de la Torre [64]	2010	52	53	On a 3-point scale, a score of at least 2.
	Kerlikowske [24]	2010	329	44.4	On a 3-point scale, a score of at least 2.

Table 9. Biomarker expression and ipsilateral recurrence risk following surgery for DCIS

Biomarker	First author and reference	Year	No. of patients	Treatment groups	Median follow-up time, months	Endocrine therapy use	Marker expression associated with increased risk of local-regional recurrence	Comments
Steroid Receptors								
ER								
	Ringberg [16]	2001	187	Lumpectomy with XRT – 66; Lumpectomy without XRT – 121	62	No	Yes (when combined with other biological markers)	The investigators evaluated a cell biological index (CBI-7) that included ER and PR negativity, overexpression of HER2, low Bcl-2 expression, accumulation of p53, nondiploidy, and high Ki-67 expression. ER negativity combined with all those markers was a strong predictor of recurrence (RR: 1.3; 95% CI: 1.0-1.6; P=0.051).
	Provenzano [22]	2003	95 (53 cases and 42 controls)	Lumpectomy without XRT – 85; Lumpectomy with XRT – 10	101	Yes	Yes	Patients with local-regional recurrence were more likely than patients without recurrence to have ER-negative disease (62% vs. 35%; OR: 0.2; P=0.01). ER negativity was individually associated with recurrence.
	Roka [23]	2004	190	Lumpectomy without XRT – 33; Lumpectomy with XRT – 99; Mastectomy with XRT – 58	61.6	Yes	Yes	The recurrence rate was higher for ER-negative DCIS than for ER-positive DCIS (12.2% vs. 3.7%; P<0.04).
	Cornfield [46]	2004	151	All patients were treated with lumpectomy without	65	No	No	ER was not associated with disease recurrence in either univariate or multivariate analysis.

			XRT				
Barnes [35]	2005	129	Lumpectomy – 89; Mastectomy – 40 (8 patients received XRT)	Not provided	No	No	ER was not associated with disease recurrence in multivariate analysis.
Kepple [59]	2006	94	Lumpectomy without XRT – 17; Lumpectomy with XRT – 20; Mastectomy – 57	48 months	Yes	Unknown	Difficult to assess effect. Only 37 patients underwent lumpectomy, and there were only 4 recurrences in that group. Some of those 37 patients received radiotherapy and some did not.
Barnes [54]	2006	161	Lumpectomy – 103; Mastectomy – 47; Information unavailable for 11 patients. No information available on XRT	Not provided	Unknown	No	ER was not associated with disease recurrence in multivariate analysis.
Wilson [37]	2006	129	Patients underwent definitive surgery for DCIS, but no details were provided	Not provided	No	No	ER was not associated with disease recurrence in either univariate or multivariate analysis.
de Roos [43]	2007	87	Lumpectomy – 39; Mastectomy – 48 (21 patients received XRT)	49.8	No	No	ER was not associated with disease recurrence in either univariate or multivariate analysis.
Millar [31]	2007	60	Lumpectomy with or without XRT – 56 (51 received XRT); Mastectomy without XRT – 4	98	No	No	In the univariate analysis conducted with clinicopathological parameters, ER was not associated with disease recurrence.
Kulkarni [21]	2008	69	Lumpectomy without XRT – 26; Lumpectomy with XRT – 43	Mean time to recurrence: 38.5	Yes	No	The investigators concluded that the biological marker, ER, is not an independent predictor of recurrence.
Altintas [15]	2009	159	Lumpectomy – 112; Mastectomy – 45 Information unavailable for 2 patients. No information available on XRT	54	No	No	The investigators concluded that the biological marker, ER, is not an independent predictor of recurrence.
Kerlikowske [24]	2010	329 (Controls with no recurrence, 186; cases with invasive recurrence, 72; cases with DCIS recurrence, 71)	All patients were treated with lumpectomy alone	98	No	Yes (when combined with other biomarkers)	In the univariate analysis, patients with DCIS recurrence were more likely than those without recurrence to have ER-negative disease (31% vs. 20%). ER negativity was individually associated with DCIS recurrence. In addition, ER negativity combined with either HER2 positivity or Ki-67 positivity was also associated with DCIS recurrence. In the multivariate analysis, the phenotype ER-HER2 ⁺ Ki-67 ⁺ was a strong predictor of subsequent DCIS recurrence (HR: 5.8; 95% CI: 2.4-14).
Zhou [25]	2010	392	Lumpectomy without XRT – 158; Lumpectomy with XRT – 140; Mastectomy – 94	97.5	No	No	The investigators looked at basal-like tumors (tumors negative for ER, PR, and HER2). In the univariate and multivariate analyses, basal-like DCIS was associated with a higher risk of invasive recurrence than non-basal-like DCIS. However, the difference was not statistically significant.
Witkiewicz [61]	2010	97	All patients underwent lumpectomy; no information was available about XRT	110.8	No	No	The investigators did not find ER to be an independent predictor of recurrence.
Holmes [58]	2011	141	All patients underwent lumpectomy alone	125	No	No	In the univariate and multivariate analyses, ER was not a predictor of recurrence.

PR

Ringberg [16]	2001	187	Lumpectomy with XRT – 66; Lumpectomy without XRT – 121	62	No	Yes (when combined with other biological markers)	The investigators evaluated a cell biological index (CBI-7) that included ER and PR negativity, overexpression of HER2, low Bcl-2 expression, accumulation of p53, nondiploidy, and high Ki-67 expression. PR negativity combined with all those markers was a strong predictor of recurrence (RR: 1.3; 95% CI: 1.0-1.6; P=0.051).
Provenzano [22]	2003	95	Lumpectomy without XRT – 85; Lumpectomy with XRT – 10	101	Yes	Yes	Patients with local-regional recurrence were more likely than those without recurrence to have PR-negative disease (63% vs. 34%; OR: 0.2; P=0.04). PR negativity was individually associated with recurrence.
Roka [23]	2004	190	Lumpectomy without XRT – 33; Lumpectomy with XRT – 99; Mastectomy with XRT – 58	61.6	Yes	No	PR-negative DCIS was associated with a higher rate of recurrence than PR-positive DCIS (9.1% vs. 3.6%), but this difference did not reach statistical significance.
Cornfield [46]	2004	151	All patients were treated with lumpectomy alone	65	No	No	PR was not associated with disease recurrence in either univariate or multivariate analysis.
Kepple [59]	2006	94	Lumpectomy without XRT – 17; Lumpectomy with XRT – 20; Mastectomy – 57	48 months	Yes	Unknown	Difficult to assess effect. Only 37 patients underwent lumpectomy, and there were only 4 recurrences in that group. Some of those 37 patients received radiotherapy and some did not.
de Roos [43]	2007	87	Lumpectomy – 39; Mastectomy – 48 (21 patients received XRT)	49.8	No	No	PR was not associated with disease recurrence in either univariate or multivariate analysis.
Millar [31]	2007	60	Lumpectomy with or without XRT – 56 (51 received XRT); Mastectomy without XRT – 4	98	No	No	In the univariate analysis conducted with clinicopathological parameters, PR was not associated with disease recurrence.
Kulkarni [21]	2008	69	Lumpectomy without XRT – 26; Lumpectomy with XRT – 43	Mean time to recurrence: 38.5	Yes	No	The investigators concluded that the biological marker, PR, is not an independent predictor of recurrence.
Altintas [15]	2009	159	Lumpectomy – 112; mastectomy – 45 Information unavailable for 2 patients. No information available on XRT	54	No	No	The investigators concluded that the biological marker, PR, is not an independent predictor of recurrence.
Zhou [25]	2010	392	Lumpectomy without XRT – 158; Lumpectomy with XRT – 140; Mastectomy – 94	97.5	No	No	The investigators looked at basal-like tumors (tumors negative for ER, PR, and HER2). In the univariate and multivariate analyses, basal-like DCIS was associated with a higher risk of local recurrence (HR: 1.7) than non-basal-like DCIS (HR: 1.8). However, the difference was not statistically significant.
Kerlikowske [24]	2010	329 (Controls with no recurrence, 186; cases with invasive recurrence, 72; cases with DCIS recurrence, 71)	All patients were treated with lumpectomy alone	98	No	No	PR was not associated with invasive or DCIS recurrence in either univariate or multivariate analysis.
Witkiewicz [61]	2010	97	All patients underwent lumpectomy (no information was	110.8	No	No	The investigators did not find PR to be an independent predictor of recurrence.

				available about XRT)					
	Holmes [58]	2011	141	All patients underwent lumpectomy alone	125	No	No		PR was not associated with disease recurrence in either univariate or multivariate analysis.
AR	Provenzano [22]	2003	95	Lumpectomy without XRT—85; Lumpectomy with XRT—10	101	Yes	No		The investigators did not find AR to be associated with disease recurrence.
Proliferation marker Ki-67									
	Ringberg [16]	2001	187	Lumpectomy with XRT—66; Lumpectomy without XRT—121	62	No	Yes (when combined with other biological markers)		The investigators evaluated a cell biological index (CBI-7) that included ER and PR negativity, overexpression of HER2, low Bcl-2 expression, accumulation of p53, nondiploidy, and high Ki-67 expression. High Ki-67 expression combined with all those markers was a strong predictor of recurrence (RR: 1.3; 95% CI: 1.0-1.6; $P=0.051$).
	Chasle [38]	2003	50	All patients underwent lumpectomy followed by XRT	Unknown	No	Yes (when combined with cyclin A)		A global proliferation factor (GPF) was calculated that was a sum of Ki-67 and cyclin A. In both univariate and multivariate analyses, GPF was an independent predictor of recurrence.
	Cornfield [46]	2004	151	All patients were treated with lumpectomy alone	65	No	No		Ki-67 was not associated with disease recurrence in either univariate or multivariate analysis.
	Barnes [35]	2005	129	Lumpectomy—89; Mastectomy—40 (8 patients received XRT)	Not provided	No	Yes		Patients with recurrence were more likely than patients without recurrence to have high proliferative activity (15.5% vs. 10.9%; $P=0.005$). In the multivariate analysis, Ki-67 was an independent predictor of recurrence (OR: 1.03, 95% CI: 1.00-1.06; $P=0.038$)
	Wilson [37]	2006	129	Patients underwent definitive surgery for DCIS, but no details were provided	Not provided	No	No		In the univariate analysis, patients with recurrence were more likely than patients without recurrence to have high proliferative activity compared to patients without a recurrence (71.4 vs. 42.2%, $P=0.006$). However, in the multivariate analysis, Ki-67 was not an independent predictor of recurrence.
	Barnes [54]	2006	161	Lumpectomy—103; Mastectomy—47; Information unavailable for 11 patients. No information available on XRT	Not provided	Unknown	Yes		In the multivariate analysis, Ki-67 was an independent predictor of recurrence (OR: 1.03, 95% CI: 1.00-1.06; $P=0.006$).
	Gauthier [45]	2007	70	Patients underwent definitive surgery for DCIS, but no details were provided	Not provided	Unknown	Yes (as an independent factor and combined with p16 expression and COX-2 expression)		High Ki-67 expression was an independent predictor of recurrence (HR: 2.7, 95% CI: 1.2-5.9). In addition, patients with recurrence were more likely than patients without recurrence to have the combination of high Ki-67, high p16, and high COX-2 expression.
	Altintas [15]	2009	159	Lumpectomy—112; mastectomy—45 Information unavailable for 2 patients. No information available on XRT	54	No	No		The investigators concluded that the biological marker, Ki67, is not an independent predictor of recurrence.
	Kerlikowske [24]	2010	329 (Controls with no recurrence, 186;	All patients were treated with lumpectomy alone	98	No	Yes		In the univariate analysis, patients with invasive recurrence were more likely than those without recurrence to exhibit the pheno-

cases with invasive recurrence, 72; cases with DCIS recurrence, 71)

type p16⁺COX-2⁺Ki-67⁺ or p16⁺Ki-67⁺. Ki-67 was individually associated with DCIS recurrence. In addition, patients with DCIS recurrence were more likely to have ER-Ki-67⁺ or p16⁺COX-2-Ki-67⁺ disease than were patients without recurrence. In the multivariate analysis, the phenotype p16⁺COX-2⁺Ki-67⁺ was a strong predictor of invasive recurrence (HR: 2.2; 95% CI: 1.1-4.5). Phenotypes p16⁺COX-2⁺Ki-67⁺ (HR: 3.7; 95% CI: 1.7-7.9) and ER-HER2⁺Ki-67⁺ (HR: 5.8, 95% CI: 2.4-14) were strong predictors of DCIS recurrence.

Cell Cycle Regulation and Apoptotic Markers

cyclin D1

Jirström [42]	2003	177	Lumpectomy without XRT – 64; Lumpectomy with XRT – 113	63	No	Yes	The investigators reported cyclin D1 expression to be strongly and inversely related with risk of ipsilateral local recurrence.
Chasle [38]	2003	50	All patients underwent lumpectomy followed by XRT	Unknown	No	No	Cyclin D1 was not a predictor of recurrence in either univariate or multivariate analysis.
de Roos [43]	2007	87	Lumpectomy – 39; Mastectomy – 48 (21 patients received XRT)	49.8	No	No	Cyclin D1 was not associated with disease recurrence in either univariate or multivariate analysis.
Millar [31]	2007	60	Lumpectomy with or without XRT – 56 (51 received XRT); Mastectomy without XRT – 4	98	No	No	In the univariate analysis conducted with clinicopathological parameters, cyclin D1 was not associated with disease recurrence.
Kulkarni [21]	2008	69	Lumpectomy without XRT – 26; Lumpectomy with XRT – 43	Mean time to recurrence: 38.5	Yes	No	The investigators concluded that the biological marker, cyclin D1, is not an independent predictor of recurrence.

cyclin A

Chasle [38]	2003	50	All patients underwent lumpectomy followed by XRT	Unknown	No	Yes (when combined with Ki-67)	A global proliferation factor (GPF) was calculated that was a sum of Ki-67 and cyclin A. In the univariate and multivariate analyses, GPF was an independent predictor of recurrence.
Millar [31]	2007	60	Lumpectomy with or without XRT – 56 (51 received XRT); Mastectomy without XRT – 4	98	No	No	In the univariate analysis conducted with clinicopathological parameters, cyclin A was not associated with disease recurrence.

cyclin E

Jirström [42]	2003	177	Lumpectomy without XRT – 64; Lumpectomy with XRT – 113	63	No	No	The investigators concluded that the biological marker, cyclin E, is not an independent risk factor for recurrence.
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p16

Jirström [42]	2003	177	Lumpectomy without XRT – 64; Lumpectomy with XRT – 113	63	No	No	The investigators concluded that the biological marker, p16, is not an independent risk factor for recurrence.
Gauthier [45]	2007	70	Patients underwent definitive surgery for DCIS, but no details were provided	Not provided	Unknown	Yes (when combined with high COX-2 expression and high Ki-67 expression)	Patients with recurrence were more likely than patients without recurrence to have the combination of high Ki-67, high p16, and high COX-2 expression.
Kerlikowske [24]	2010	329 (Controls with no recurrence, 186; cases with	All patients were treated with lumpectomy alone	98	No	Yes	In the univariate analysis, patients with invasive recurrence were more likely than those without recurrence to have p16-positive disease (57% vs. 30%). p16 positiv-

								invasive recurrence, 72; cases with DCIS recurrence, 71)329 patients
								ity combined with Ki-67 positivity and COX-2 positivity was also associated with invasive recurrence. p16 was individually associated with invasive recurrence. In addition, p16 positivity combined with Ki-67 positivity and COX-2 negativity was associated with DCIS recurrence. In the multivariate analysis, the phenotype p16 ⁺ COX-2 ⁺ Ki-67 ⁺ was a strong predictor of invasive recurrence (HR: 2.2; 95% CI: 1.1-4.5). Also, the phenotype p16 ⁺ COX-2 ⁻ Ki-67 ⁺ was a strong predictor of DCIS recurrence (HR: 3.7; 95% CI: 2.4-14).
p21								
	Provenzano [22]	2003	95	Lumpectomy without XRT—85; Lumpectomy with XRT—10	101	Yes	Yes	Patients with local-regional recurrence were more likely than those without recurrence to have p21-positive disease (54% vs. 15%; OR: 6.0; <i>P</i> =0.01). p21 positivity was individually associated with recurrence. According to the multiple conditional logistic regression analysis, p21 expression was an independent predictor of recurrence (OR range: 4.31–6.54).
	Chasle [38]	2003	50	All patients underwent lumpectomy followed by XRT	Unknown	No	No	According to univariate and multivariate analyses, p21 was not observed to be an independent predictor of recurrence.
	Cornfield [46]	2004	151	All patients were treated with lumpectomy alone	65	No	No	p21 was not associated with disease recurrence in either univariate or multivariate analysis.
	Kulkarni [21]	2008	69	Lumpectomy without XRT—26; Lumpectomy with XRT—43	Mean time to recurrence: 38.5	Yes	No	The investigators concluded that the biological marker, p21, is not an independent predictor of recurrence.
p27								
	Millar [31]	2007	60	Lumpectomy with or without XRT—56 (51 received XRT); Mastectomy without XRT—4	98	No	No	In the univariate analysis conducted with clinicopathological parameters, p27 was not associated with disease recurrence.
	Jirström [42]	2003	177	Lumpectomy without XRT—64; Lumpectomy with XRT—113	63	No	No	The investigators concluded that the biological marker, p27, is not an independent risk factor for recurrence.
p53								
	Hieken [48]	2001	103	Lumpectomy without XRT—34; Lumpectomy with XRT—41; Mastectomy—28	58 (mean follow-up time)	Yes	Yes	p53 was expressed in 63% of patients with recurrence and 24% of patients without recurrence (<i>P</i> =0.03). The investigators concluded that strong p53 expression is associated with ipsilateral tumor recurrence.
	Ringberg [16]	2001	187	Lumpectomy with XRT—66; Lumpectomy without XRT—121	62	No	Yes (when combined with other biological markers)	The investigators evaluated a cell biological index (CBI-7) that included ER and PR negativity, overexpression of HER2, low Bcl-2 expression, accumulation of p53, nondiploidy, and high Ki-67 expression. High p53 expression combined with all those markers was a strong predictor of recurrence (RR: 1.3; 95% CI: 1.0-1.6; <i>P</i> =0.051).
	Provenzano [22]	2003	95 (53 cases and 42 controls)	Lumpectomy without XRT—85; Lumpectomy with XRT—10	101	Yes	No	The investigators did not find p53 to be associated with disease recurrence.
	Chasle [38]	2003	50	All patients underwent lumpectomy followed by XRT	Unknown	No	No	p53 was not an independent predictor of recurrence in either univariate or multivariate analysis.

	Cornfield [46]	2004	151	All patients were treated with lumpectomy without XRT	65	No	No	p53 was not associated with disease recurrence in either univariate or multivariate analysis.
	Roka [23]	2004	190	Lumpectomy without XRT –33; Lumpectomy with XRT –99; Mastectomy and XRT –58	61.6	Yes	No	The investigators did not find p53 to be an independent predictor of disease recurrence.
	Kepple [59]	2006	94	Lumpectomy without XRT –17; Lumpectomy with XRT –20; Mastectomy –57	48 months	Yes	Unknown	Difficult to assess effect. Only 37 patients underwent lumpectomy, and there were only 4 recurrences in that group. Some of those 37 patients received radiotherapy and some did not.
	de Roos [43]	2007	87	Lumpectomy –39; Mastectomy –48 (21 patients received XRT)	49.8	No	Yes	p53 was an independent predictor of disease recurrence in multivariate (HR: 3.0, 95% CI: 1.1-8.2, $P=0.036$) and univariate (HR: 3.5, 95% CI: 1.3-9.3, $P=0.014$) analyses.
	Kulkarni [21]	2008	69	Lumpectomy without XRT –26; Lumpectomy with XRT –43	Mean time to recurrence: 38.5	Yes	No	The investigators concluded that the biological marker, p53, is not an independent predictor of recurrence.
	Kerlikowske [24]	2010	329 (Controls with no recurrence, 186; cases with invasive recurrence, 72; cases with DCIS recurrence, 71)	All patients were treated with lumpectomy alone	98	No	No	p53 was not associated with invasive or DCIS recurrence either individually or when combined with other phenotypes.
Bcl-2	Ringberg [16]	2001	187	Lumpectomy with XRT –66; Lumpectomy without XRT –121	62	No	Yes (when combined with other biological markers)	The investigators evaluated a cell biological index (CBI-7) that included ER and PR negativity, overexpression of HER2, low Bcl-2 expression, accumulation of p53, nondiploidy, and high Ki-67 expression. Low Bcl-2 expression combined with all those markers was a strong predictor of recurrence (RR: 1.3; 95% CI: 1.0-1.6; $P=0.051$).
	Provenzano [22]	2003	95	Lumpectomy without XRT –85; Lumpectomy with XRT –10	101	Yes	Yes	Patients with local-regional recurrence were more likely than those without recurrence to have Bcl-2-negative disease (66% vs. 26%; $P=0.003$; OR: 0.18).
	Cornfield [46]	2004	151	All patients were treated with lumpectomy alone	65	No	No	Bcl-2 was not associated with disease recurrence in either univariate or multivariate analysis.
Survivin	Barnes [54]	2006	161	Lumpectomy –103; Mastectomy –47; Information unavailable for 11 patients. No information available on XRT	Not provided	Unknown	Yes (not as an independent factor, but when combined with COX-2 expression)	Patients with recurrence were more likely than those without recurrence to have co-expression of COX-2 and cytoplasmic survivin (70% vs. 41%; $P=0.013$).
c-myc	Altintas [15]	2009	159	Lumpectomy –112; Mastectomy –45 Information unavailable for 2 patients. No information available on XRT	54	No	No	The investigators concluded that the biological marker, c-myc, is not an independent predictor of recurrence.
Angiogenesis-related proteins								
VEGF								
	Hieken [48]	2001	103	Lumpectomy	58	Yes	No	The investigators did not find

				without XRT – 34; Lumpectomy with XRT – 41; Mastec- tomy – 28	(mean fol- low-up time)			VEGF to be an independent pre- dictor of disease recurrence.
Epidermal Growth Factor Receptor Family								
HER1								
	Barnes [35]	2005	129	Lumpectomy – 89; Mastectomy – 40 (8 patients received XRT)	5 years	Unknown	No	The investigators concluded that the biological marker, HER1, is not an independent predictor of re- currence.
	Altintas [15]	2009	159	Lumpectomy – 112; mastectomy – 45 Information una- vailable for 2 pa- tients. No infor- mation on XRT	54	No	No	The investigators concluded that the biological marker, HER1, is not an independent predictor of re- currence.
HER2								
	Ringberg [16]	2001	187	Lumpectomy with XRT – 66; Lumpec- tomy without XRT – 121	62	No	Yes (when combined with other biological markers)	The investigators evaluated a cell biological index (CBI-7) that in- cluded ER and PR negativity, overexpression of HER2, low Bcl-2 expression, accumulation of p53, nondiploidy, and high Ki-67 ex- pression. HER2 positivity combi- ned with all those markers was a strong predictor of recurrence (RR: 1.3; 95% CI: 1.0-1.6; P=0.051).
	Provenzano [22]	2003	95	Lumpectomy without XRT – 85; Lumpectomy with XRT – 10	101	Yes	Yes	Patients with local-regional re- currence were more likely than those without recurrence to have HER2-positive disease (41% vs. 12%; OR: 5.0; P=0.008).
	Roka [23]	2004	190	Lumpectomy without XRT – 33; Lumpectomy with XRT – 99; Mastec- tomy with XRT – 58	61.6	Yes	No	The investigators did not find HER2 to be an independent pre- dictor of disease recurrence.
	Cornfield [46]	2004	151	All patients were treated with lum- pectomy alone	65	No	No	HER2 was not associated with disease recurrence in either uni- variate or multivariate analysis.
	Barnes [35]	2005	129	Lumpectomy – 89; Mastectomy – 40 (8 patients received XRT)	5 years	Unknown	No	The investigators concluded that the biological marker, HER2, is not an independent predictor of re- currence.
	Barnes [54]	2006	161	Lumpectomy – 103; Mastectomy – 47; Information una- vailable for 11 pa- tients. No infor- mation available on XRT	Not pro- vided	Unknown	No	HER2 was not associated with disease recurrence in multivariate analysis.
	Kepple [59]	2006	94	Lumpectomy without XRT – 17; Lumpectomy with XRT – 20; Mastec- tomy – 57	48 months	Yes	Unknown	Difficult to assess effect. Only 37 patients underwent lumpectomy, and there were only 4 recurrences in that group. Some of those 37 patients received radiotherapy and some did not.
	de Roos [43]	2007	87	Lumpectomy – 39; Mastectomy – 48 (21 patients re- ceived XRT)	49.8	No	No	HER2 overexpression was associ- ated with recurrence in univariate analysis (HR: 3.1, 95% CI: 1.1-8.7; P=0.032). However, multivariate analysis did not show HER2 overexpression to be an inde- pendent predictor of recurrence.
	Kulkarni [21]	2008	69	Lumpectomy without XRT – 26; Lumpectomy with XRT – 43	Mean time to recur- rence: 38.5	Yes	No	The investigators concluded that the biological marker, HER2, is not an independent predictor of re- currence.
	Altintas [15]	2009	159	Lumpectomy – 112; mastectomy – 45 Information una- vailable for 2 pa- tients. No infor- mation available on	54	No	No	The investigators concluded that the biological marker, HER2, is not an independent predictor of re- currence.

				XRT				
	Stackiewicz [74]	2010	84	Lumpectomy—80 (43 patients received XRT); Mastectomy—4	94.8	Yes	No	The investigators concluded that the biological marker, HER2, is not an independent risk factor for recurrence.
	Kerlikowske [24]	2010	329 (Controls with no recurrence, 186; cases with invasive recurrence, 72; cases with DCIS recurrence, 71)	All patients were treated with lumpectomy alone	98	No	Yes	In the univariate analysis, patients with DCIS recurrence were more likely than patients without recurrence to have HER2-positive disease (30% vs. 13%). HER2 was individually associated with DCIS recurrence. Also, patients with DCIS recurrence were more likely than patients without recurrence to exhibit the ER-HER2 ⁺ phenotype (19% vs. 6.4%). In addition, a multivariate analysis showed that the phenotype ER-HER2 ⁺ Ki-67 ⁺ was a strong predictor of subsequent DCIS recurrence (OR: 5.8; 95% CI: 2.4-14).
	Zhou [25]	2010	392	Lumpectomy without XRT—158; Lumpectomy with XRT—140; Mastectomy—94	97.5	No	No	The investigators looked at basal-like tumors (tumors negative for ER, PR, and HER2). In the univariate and multivariate analyses, basal-like DCIS was associated with a higher risk of local recurrence (HR: 1.7) than non-basal-like DCIS (HR: 1.8). However, the difference was not statistically significant. (Note: The authors do not state in the paper how many patients with basal-like DCIS developed a recurrence—they only report HRs in the tables.)
	Witkiewicz [61]	2010	97	All patients underwent lumpectomy (no information was available about XRT)	110.8	No	No	The investigators did not find HER2 to be an independent predictor of recurrence.
	Holmes [58]	2011	141	All patients underwent lumpectomy alone	125	No	Yes	Univariate analysis with respect to time to recurrence found HER2 overexpression to be associated with local recurrence ($P=0.028$). In the multivariate analysis, HER2 overexpression was an independent predictor of disease recurrence (HR: 1.82, 95% CI: 1.03-3.22, $P=0.041$).
HER3	Barnes [35]	2005	129	Lumpectomy—89; Mastectomy—40 (8 patients received XRT)	5 years	Unknown	No	The investigators concluded that the biological marker, HER3, is not an independent predictor of recurrence.
	Altintas [15]	2009	159	Lumpectomy—112; mastectomy—45 Information unavailable for 2 patients. No information on XRT	54	No	No	The investigators concluded that the biological marker, HER3, is not an independent predictor of recurrence.
HER4	Barnes [35]	2005	129	Lumpectomy—89; Mastectomy—40 (8 patients received XRT)	5 years	Unknown	Yes	The investigators concluded that HER4 expression is an independent predictor of a reduced risk of recurrence (OR: 0.69, 95% CI: 0.48-0.98, $P=0.038$).
	Altintas [15]	2009	159	Lumpectomy—112; mastectomy—45 Information unavailable for 2 patients. No information on XRT	54	No	No	The investigators concluded that the biological marker, HER4, is not an independent predictor of recurrence.
<i>Extracellular matrix-related proteins</i>								
CD10								
	Toussaint	2010	154	Surgical infor-	6 years	Yes	Yes	According to the multivariate

	[60]							<p>mation was reported according to VNPI. 58% of patients with low VNPI were treated with lumpectomy alone; 41% of those with intermediate VNPI received XRT following lumpectomy; 81% of patients with high VNPI underwent mastectomy</p>					<p>analysis, CD10 was an independent predictor of recurrence. Patients with low CD10 expression were more likely than those with high CD10 expression to develop recurrence (HR: 2.39, 95% CI: 1.52-3.76, $P=0.001$).</p>
	Witkiewicz [61]	2010	97					<p>All patients underwent lumpectomy (no information was available about XRT)</p>	110.8	No	Yes		<p>In the multivariate analysis, CD10 was an independent predictor of recurrence. Patients with recurrence were more likely than those without recurrence to exhibit strong stromal CD10 expression (OR: 10.2, 95% CI: 2.7, 37.7).</p>
SPARC	Witkiewicz [61]	2010	97					<p>All patients underwent lumpectomy (no information was available about XRT)</p>	110.8	No	Yes		<p>In the multivariate analysis, SPARC was an independent predictor of recurrence. Patients with recurrence were more likely than those without recurrence to exhibit strong stromal SPARC expression (OR: 3.9, 95% CI: 1.1, 14.3).</p>
COX-2	Barnes [54]	2006	161					<p>Lumpectomy – 103; Mastectomy – 47; Information unavailable for 11 patients. No information available on XRT</p>	Not provided	Unknown	Yes (as an independent factor and in combination with survivin)		<p>In the multivariate analysis, COX-2 was an independent predictor of recurrence. In addition, patients with recurrence were more likely than those without recurrence to have co-expression of COX-2 and cytoplasmic survivin compared to patients without recurrences co-expressing both proteins (70% vs. 41%; $P=0.013$).</p>
	Gauthier [45]	2007	70					<p>Patients underwent definitive surgery for DCIS, but no details were provided</p>	Not provided	Unknown	Yes (when combined with high p16 expression and high Ki-67 expression)		<p>COX-2 was not an independent predictor of recurrence. However, patients with recurrence were more likely than those without recurrence to express the combination of high Ki-67, high p16, and high COX-2 expression.</p>
	Kulkarni [21]	2008	69					<p>Lumpectomy without XRT – 26; Lumpectomy with XRT – 43</p>	Mean time to recurrence: 38.5	Yes	Yes		<p>In the multivariate analysis, COX-2 expression was significantly associated with increased risk of recurrence (OR: 7.89; 95% CI 1.7-36.2).</p>
	Kerlikowske [24]	2010	329					<p>All patients were treated with lumpectomy alone</p>	98	No	Yes (in combination with other markers)		<p>In the univariate analysis, patients with invasive recurrence were more likely than those without recurrence to exhibit the phenotype p16⁺COX-2⁺Ki-67⁺ (23% vs. 8.5%). Patients with DCIS recurrence were more likely than those without a recurrence to exhibit the phenotype p16⁺COX-2⁺Ki-67⁺ (19% vs. 2.6%). COX-2 was not individually associated with recurrence. In the multivariate analysis the phenotype p16⁺COX-2⁺Ki-67⁺ was a strong predictor of invasive recurrence (HR: 2.2; 95% CI: 1.1-4.5). Another phenotype, p16⁺COX-2⁺Ki-67⁺, was a strong predictor of DCIS recurrence (HR: 3.7; 95% CI: 1.7-7.9).</p>

XRT, radiotherapy; VNPI, Van Nuys Prognostic Index.