


Short Research Communication

# MicroRNA-210 and Endoplasmic Reticulum Chaperones in the Regulation of Chemoresistance in Glioblastoma

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## Abstract

Glioblastoma multiforme (GBM) is the commonest primary brain tumour in adults characterized by relentless recurrence due to resistance towards the standard chemotherapeutic agent temozolomide (TMZ). Prolyl 4-hydroxylase, beta polypeptide (P4HB), an endoplasmic reticulum (ER) chaperone, is known to be upregulated in TMZ-resistant GBM cells. MicroRNAs (miRNAs) are non-protein-coding transcripts that may play important roles in GBM chemoresistance. We surmised that miRNA dysregulations may contribute to P4HB upregulation, hence chemoresistance. We found that miRNA-210 (miR-210) was P4HB-targeting and was highly downregulated in TMZ-resistant GBM cells. Forced overexpression of miR-210 led to P4HB downregulation and a reduction in TMZ-resistance. A reciprocal relationship between their expressions was also verified in clinical glioma specimens. Our study is the first to demonstrate a potential link between miR-210 and ER chaperone in determining chemosensitivity in GBM. The findings have important translational implications in suggesting new directions of future studies.

Key words: Glioblastoma, miRNA, P4HB, ER stress response, chemoresistance, temozolomide

## Introduction

Glioblastoma multiforme (GBM), a World Health Organization (WHO) Grade IV glioma, is the commonest and most malignant form of primary brain tumour [1-3]. GBM is highly invasive and refractory towards the standard chemotherapeutic agent temozolomide (TMZ), resulting in high recurrence and mortality rates [4].

Prolyl 4-hydroxylase, beta polypeptide (P4HB) is a chaperone protein of the endoplasmic reticulum stress response (ERSR) pathway. While its normal function is to prevent unfolded protein aggregation, P4HB has also been found to prevent chemotherapeutics-induced apoptosis in cancer cells [5, 6]. In-

deed, our group have previously identified P4HB to be highly expressed in TMZ-resistant GBM cells [7]. This conferred TMZ-resistance by suppressing the activation of the downstream ERSR pathway that would otherwise induce apoptosis [8].

MicroRNAs (miRNAs) are small, non-coding RNAs about 22 nucleotides long. They regulate gene expression post-transcriptionally by binding to the 3' untranslated region (3' UTR) of the target genes and cause messenger RNA (mRNA) degradation and/or translation inhibition [9, 10]. Dysregulated miRNA expressions are found in various cancers and are intimately involved in cancer progression [11]. Hypoxia

is an important inducer of ERSR in GBM and is also commonly associated with miRNA dysregulation. For instance, miR-210 is consistently upregulated under hypoxia and highly dysregulated in GBM, but its role in GBM chemoresistance has not been studied [12, 13]. Interestingly, miR-210 is also known to target P4HB [14]. We therefore conducted the present study to investigate whether and how miR-210 may interact with the ERSR pathway, in particular its chaperone protein P4HB, in affecting GBM response to TMZ. The hypothesis was that miR-210 would be downregulated in TMZ-resistant cells and that its upregulation would improve chemoresponsiveness.

## Methods and Materials

### GBM cell culture and TMZ-resistant subclones

Two human GBM cell lines were used: D54-MG (a gift from Duke University Medical Center, Durham, NC, USA) and U87-MG (American Type Culture Collection, Manassas, VA, USA). D54-MG cells were cultured in Dulbecco's Modified Eagle's Medium (DMEM)/F-12, and U87-MG in Minimum Essential Medium (MEM)  $\alpha$ , all of which were supplemented with 10% fetal bovine serum (FBS) and 1% Penicillin-Streptomycin (PS) (all from Gibco® by Life Technologies, Grand Island, NY, USA). Both cell lines were cultured in a 37°C humidified incubator with 5% CO<sub>2</sub>. Isogenic TMZ-resistant subclones D54-R and U87-R were derived after chronic exposure of the parental cells D54-S and U87-S to TMZ (Temodal; Schering-Plough Corporation, Kenilworth, NJ, USA) as previously described [7].

### Clinical glioma tumour samples

Clinical tumour samples were collected from patients who had undergone initial surgical resection before chemo-irradiation and adjuvant TMZ treatment. Tissues of WHO grade II to IV, confirmed by a neuropathologist, were used. The study was approved by the Institution Review Board of our institution and signed consent forms by patients or their legal guardians were obtained.

### Quantitative real-time polymerase chain reaction (qPCR)

Relative expression levels of miR-210 and P4HB *in vitro* and in clinical specimens were quantified by qPCR. MiRNA and total RNA were extracted using the TRIzol® Reagent (Ambion® by Life Technologies, Austin, TX, USA). Purified miRNA and RNA samples were reverse-transcribed using miScript® III RT Kit (Qiagen, Venlo, Limburg, Netherlands) before qPCR amplification using miScript SYBR® Green PCR Kit (Qiagen) with the 7900HT Fast Real-Time PCR System (Applied Biosystems, Inc., Foster City, CA, USA).

MiRNA and mRNA expression levels were normalized to U6 small nuclear RNA (snRNA) and GAPDH as internal controls, respectively, and were expressed as relative fold ratios.

### MiR-210 overexpression

Transient transfections to achieve miR-210 overexpression were conducted on 6-well plates at an initial density of  $2.5 \times 10^5$  cells per well. At near confluence, cells were treated with Lipofectamine® RNAiMAX Reagent diluted with either mirVana™ miRNA mimic (which overexpresses miR-210) or mirVana™ miRNA mimic Negative Control (as negative control) (all from Ambion®).

### Cell Proliferation Assay

Cell proliferation was measured using the 3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay (Sigma-Aldrich Corporation, St Louis, MO, USA). Cells were seeded onto 96-well plates at an initial density of 5000 cells per well and were cultured for 24 hours. Culture media were changed serially to reach various concentrations of TMZ (i.e. 0, 62.5, 125, 250, 500, 1000, 2000, and 3000  $\mu$ M). Cells were collected and the optical densities (OD) were measured at an absorbance wavelength of 595 nm. The amount of cells viable relative to controls was calculated and the half-maximal inhibitory concentrations (IC<sub>50</sub>) were derived. Experiments were performed in triplicates.

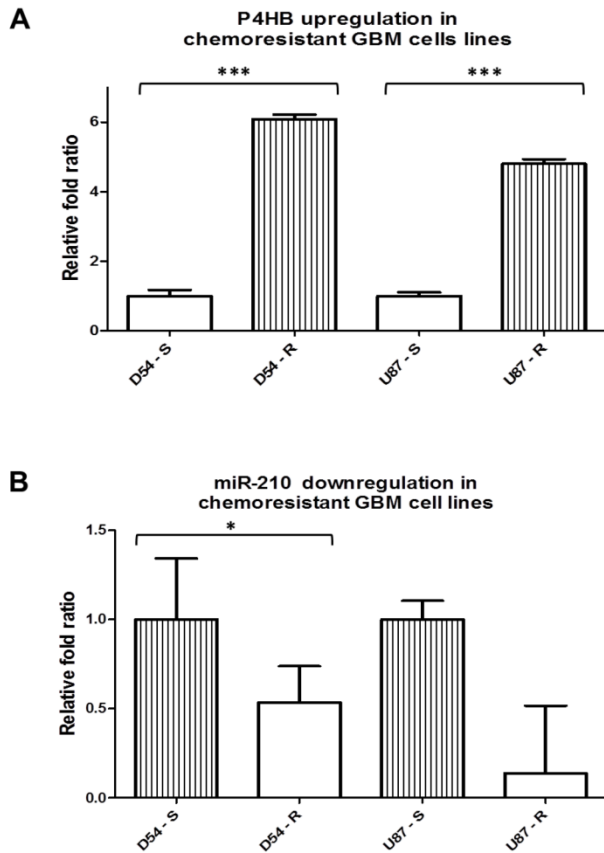
### Statistical analysis

Data analysis was performed using the Prism (version 6.0) software (Graphpad Software, Inc., La Jolla, CA, USA). Statistical significance was determined using student's *t*-test and *p*-value < 0.05 was considered statistically significant.

## Results

### P4HB is upregulated and miR-210 is downregulated in chemoresistant GBM

To verify the involvement of P4HB in GBM chemoresistance, and that miR-210 would be its potential regulator, their intrinsic expressions were quantified. As expected, P4HB expression was upregulated in the TMZ-resistant cells D54-R and U87-R by approximately 6-folds and 5-folds, respectively, when compared with the parental cells D54-S and U87-S (Fig. 1A). Conversely, the TMZ-resistant D54-R and U87-R cells expressed lower levels of miR-210 in comparison to their TMZ-sensitive counterparts by 2-folds and 10-folds, respectively, suggesting that P4HB may potentially be under the regulation of miR-210 (Fig. 1B).



**Figure 1. P4HB is upregulated and miR-210 is downregulated in TMZ-resistant GBM cell lines. (A)** Upregulation in the relative P4HB expression levels in TMZ-resistant GBM cell lines D54-R and U87-R when compared with those in the TMZ-sensitive D54-S and U87-S cells on qPCR. **(B)** Downregulations in miR-210 expression levels in D54-R and U87-R relative to those in D54-S and U87-S cells. (\* =  $p < 0.05$ ; \*\*\* =  $p < 0.001$ )

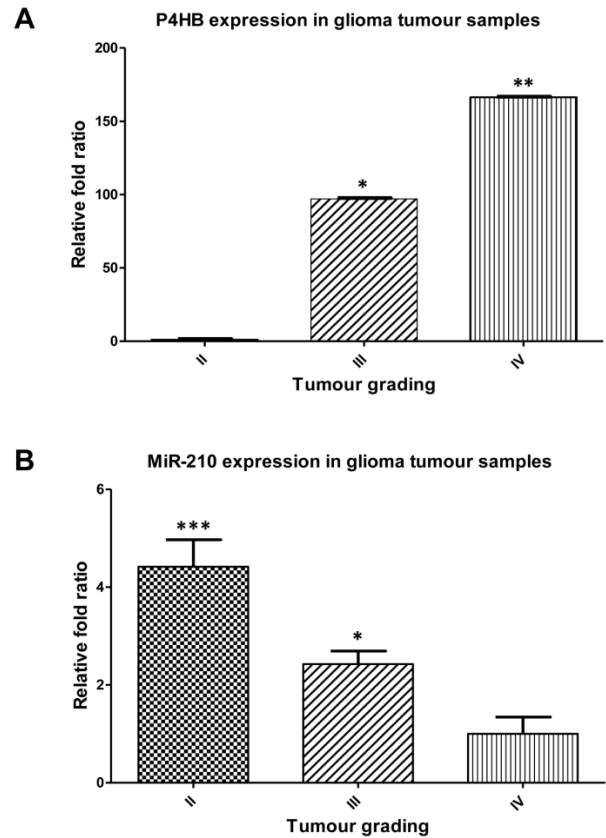
### P4HB and miR-210 expression levels in clinical glioma tissues

P4HB expressions were positively correlated with increasing tumour grades. P4HB expression was the highest in grade IV glioma at approximately 150-folds, followed by grade III at 100-folds, when compared with grade II tumours (Fig. 2A). In contrast, miR-210 expressions were negatively correlated with that of P4HB in clinical glioma tumour samples. MiR-210 was the highest in grade II gliomas at approximately 4.5-folds, followed by grade III at approximately 2.4-folds, when compared with grade IV gliomas (Fig. 2B).

### MiR-210 overexpression downregulates P4HB in GBM cell lines

We then overexpressed miR-210 to evaluate its potential regulatory action on P4HB. The transfection efficiency of the miR-210 mimic was confirmed by qPCR, which showed an increase in relative fold ratios by 100 times when compared with control (Fig. 3A). Due to the intrinsically higher expression of

miR-210 in the TMZ-sensitive cells, the absolute levels of miR-210 expression were higher in the parental D54-S and U87-S cells when compared to their resistant counterparts.



**Figure 2. P4HB is upregulated and miR-210 is downregulated in higher grade clinical tumour samples. (A)** Upregulation in the relative P4HB expression levels in grades III and IV glioma tumour samples when compared to those of grade II on qPCR. (\* =  $p < 0.05$ ; \*\* =  $p < 0.01$  compared to grade II, respectively) **(B)** Downregulations in miR-210 expression levels in grades III and IV glioma tumour samples when compared to those of grade II glioma tumour sample. (\* =  $p < 0.05$ ; \*\*\* =  $p < 0.001$  compared to grade IV, respectively)

P4HB expressions at the mRNA levels were then determined after miR-210 overexpression. The latter caused P4HB downregulation by at least 2-folds when compared with negative controls in all four cell lines (Fig. 3B). Of note was that the absolute levels of P4HB expressions in the TMZ-resistant cell lines D54-R and U87-R remained higher in comparison to those of their respective TMZ-sensitive lines, due to the intrinsically higher expression in the former group [7].

### MiR-210 overexpression reduces chemoresistance in GBM cell lines

With its effect of downregulating P4HB expression, we then examined whether miR-210 overexpression would diminish TMZ-resistance using the

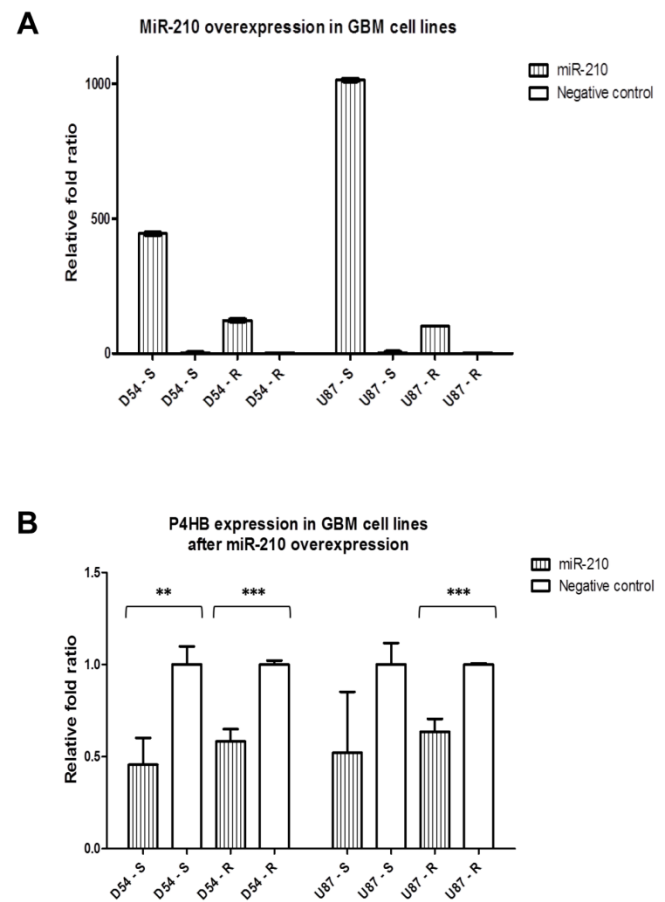
MTT cell proliferation assay (Fig. 4). Except for U87-R, the  $IC_{50}$  of TMZ for cells transfected with negative controls was around 2000  $\mu$ M. With miR-210 overexpression, the  $IC_{50}$  was reduced to about 200  $\mu$ M and 500  $\mu$ M for the D54-S and U87-S cells, respectively, and at about 500  $\mu$ M and 600  $\mu$ M for the TMZ-resistant D54-R and U87-R cells, respectively. The overall findings were consistent with the suggestion that P4HB would confer TMZ resistance in GBM, and that miR-210 overexpression may downregulate P4HB expression, resulting in greater response to TMZ cytotoxicity.

## Discussion

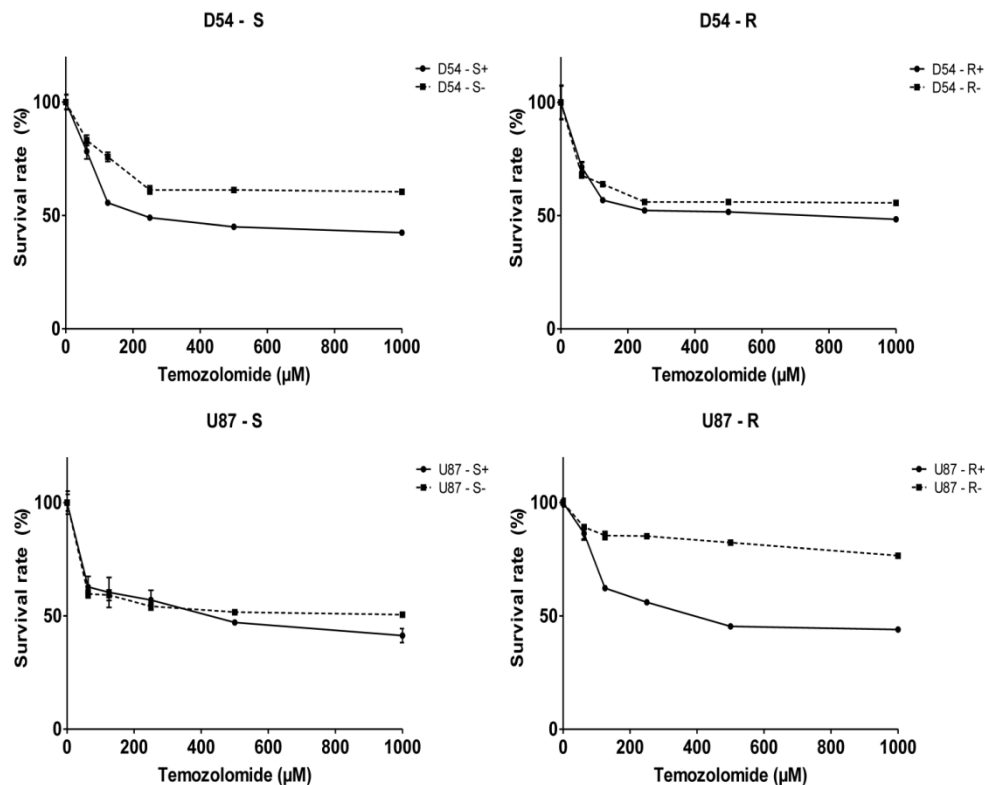
The ERSR chaperone protein, P4HB, contributes to chemoresistance in GBM cells [15]. The current study explored miRNA interference as a potential therapeutic approach to downregulate P4HB expression for the potential restoration of chemosensitivity in GBM towards TMZ. We found that miR-210 overexpression could achieve that *in vitro* in two commonly used cell lines. Interestingly, miR-210 is also widely reported to be an oncogene that is upregulated in various types of cancers, especially where intratumoural hypoxia is common [16]. Our approach of using miR-210 overexpression to improve chemosensitivity may therefore appear counterintuitive. It must be emphasized, however, that increased miR-210 expression was also found in non-malignant conditions, where hypoxia would be mild such as the placenta of pre-eclampsia patients. This has raised questions about the actual contributions of miR-210 towards tumour aggression, as miR-210 overexpression may simply be a consequence of tumour hypoxia [16-18]. Moreover, miR-210 has been found to be downregulated in other neoplasms such as ovarian cancer, either due to decreased expression or genomic deletions [19, 20]. Repression of tumour cell growth initiation by miR-210 in human head and neck, as well as pancreatic cell lines has also been reported, suggesting that miR-210 may potentially possess tumour suppressive characteristics as well as oncogenic effects [21].

In fact, previous studies have reported miR-210 to be associated with favourable prognostic outcomes. McCormick et al found the expression of miR-210 in clear cell renal cell cancer (CCRCC) to be associated with reduced proliferation, lower tumour grading, lower staging, and better patient survival, possibly due to the suppression of the von Hippel-Lindau (VHL) gene [22]. The initiation and progression of

cancers depends on the expression of a wide variety of genes and their involvement in different cellular pathways. Each miRNA may have many different mRNA targets, and therefore can affect cellular processes in different cell types differently. Similarly, miR-210 may target many genes that are involved in angiogenesis, cell proliferation and/or survival, and DNA damage repair [23]. To our knowledge, our study is the first to describe the interactions between the ERSR and miR-210, and the latter's potential effect on chemoresponsiveness. Although miR-210 is widely known for its oncogenic effects, our findings provide a novel direction for future investigations. The latter may focus on the use of miRNA interference in the treatment of chemoresistant GBM *in vivo*. Important considerations include the effective delivery of miR-210 inhibitor and target specificity.



**Figure 3. MiR-210 regulates P4HB expression. (A)** Transfection of miR-210 mimics successfully results in miR-210 overexpressions in all four GBM cell lines when compared with their respective negative controls on qPCR. **(B)** Downregulation in the relative P4HB expression levels occurs as the result of miR-210 overexpressions when compared to their respective negative controls. (\*\* =  $p < 0.01$ ; \*\*\* =  $p < 0.001$ )



**Figure 4. MiR-210 upregulation may reduce chemoresistance in GBM cells.** A decrease in  $IC_{50}$  of TMZ for cells with miR-210 overexpression (“+”) relative to its respective negative controls (“-“) in all four GBM cell lines used by MTT cell proliferation assay.

## Conclusions

MiR-210 is commonly dysregulated in various cancers and serves as a useful prognostic biomarker. Our study demonstrated that miR-210's inhibitory effects of P4HB could potentially reverse TMZ-resistance in GBM cells. Both ERSR and miR-210 are known to affect malignant phenotypes in GBM. Our study is the first to demonstrate a potential link between them in determining chemosensitivity in GBM. The findings have important translational implications in suggesting new directions of future studies for the treatment of chemoresistant GBM.

## Abbreviations

ERSR: endoplasmic reticulum stress response; GBM: glioblastoma multiforme;  $IC_{50}$ : half-maximum inhibitory concentration; miRNA: microRNA; P4HB: prolyl 4-hydroxylase, beta polypeptide; TMZ: temozolomide

## Competing Interests

The authors have declared that no competing interest exists.

## References

- Ohgaki H. Epidemiology of brain tumors. *Methods Mol Biol.* 2009; 472: 323-42.
- Fisher JL, Schwartzbaum JA, Wrensch M, et al. Epidemiology of brain tumors. *Neurol Clin.* 2007; 25: 867-90, vii.

- Louis DN, Ohgaki H, Wiestler OD, et al. The 2007 WHO classification of tumours of the central nervous system. *Acta Neuropathol.* 2007; 114: 97-109.
- Sun S, Lee D, Lee NP, et al. Hyperoxia resensitizes chemoresistant human glioblastoma cells to temozolomide. *J Neurooncol.* 2012; 109: 467-75.
- Ni M, Lee AS. ER chaperones in mammalian development and human diseases. *FEBS Lett.* 2007; 581: 3641-51.
- Fu Y, Li J, Lee AS. GRP78/BiP inhibits endoplasmic reticulum BIK and protects human breast cancer cells against estrogen starvation-induced apoptosis. *Cancer Res.* 2007; 67: 3734-40.
- Sun S, Wong TS, Zhang XQ, et al. Protein alterations associated with temozolomide resistance in subclones of human glioblastoma cell lines. *J Neurooncol.* 2012; 107: 89-100.
- Ahmed R, Oborski MJ, Hwang M, et al. Malignant gliomas: current perspectives in diagnosis, treatment, and early response assessment using advanced quantitative imaging methods. *Cancer Manag Res.* 2014; 6: 149-70.
- Ambros V, Bartel B, Bartel DP, et al. A uniform system for microRNA annotation. *Rna.* 2003; 9: 277-9.
- Bartel DP. MicroRNAs: genomics, biogenesis, mechanism, and function. *Cell.* 2004; 116: 281-97.
- Calin GA, Sevignani C, Dumitru CD, et al. Human microRNA genes are frequently located at fragile sites and genomic regions involved in cancers. *Proc Natl Acad Sci U S A.* 2004; 101: 2999-3004.
- Lai NS, Dong QS, Ding H, et al. MicroRNA-210 overexpression predicts poorer prognosis in glioma patients. *J Clin Neurosci.* 2014; 21: 755-60.
- Lee D, Sun S, Ho AS, et al. Hyperoxia resensitizes chemoresistant glioblastoma cells to temozolomide through unfolded protein response. *Anticancer Res.* 2014; 34: 2957-66.
- Fasanaro P, Greco S, Lorenzi M, et al. An integrated approach for experimental target identification of hypoxia-induced miR-210. *J Biol Chem.* 2009; 284: 35134-43.
- Sun S, Lee D, Ho AS, et al. Inhibition of prolyl 4-hydroxylase, beta polypeptide (P4HB) attenuates temozolomide resistance in malignant glioma via the endoplasmic reticulum stress response (ERSR) pathways. *Neuro Oncol.* 2013; 15: 562-77.
- Huang X, Le QT, Giaccia AJ. MiR-210—micromanagement of the hypoxia pathway. *Trends Mol Med.* 2010; 16: 230-7.
- Zhu XM, Han T, Sargent IL, et al. Differential expression profile of microRNAs in human placentas from preeclamptic pregnancies vs normal pregnancies. *Am J Obstet Gynecol.* 2009; 200: 661 e1-7.
- Pineles BL, Romero R, Montenegro D, et al. Distinct subsets of microRNAs are expressed differentially in the human placentas of patients with preeclampsia. *Am J Obstet Gynecol.* 2007; 196: 261 e1-6.

19. Giannakakis A, Sandaltzopoulos R, Greshock J, et al. miR-210 links hypoxia with cell cycle regulation and is deleted in human epithelial ovarian cancer. *Cancer Biol Ther.* 2008; 7: 255-64.
20. Wyman SK, Parkin RK, Mitchell PS, et al. Repertoire of microRNAs in epithelial ovarian cancer as determined by next generation sequencing of small RNA cDNA libraries. *PLoS One.* 2009; 4: e5311.
21. Huang X, Ding L, Bennewith KL, et al. Hypoxia-inducible mir-210 regulates normoxic gene expression involved in tumor initiation. *Mol Cell.* 2009; 35: 856-67.
22. McCormick RI, Blick C, Ragoussis J, et al. miR-210 is a target of hypoxia-inducible factors 1 and 2 in renal cancer, regulates ISCU and correlates with good prognosis. *Br J Cancer.* 2013; 108: 1133-42.
23. Mei Y, Gao C, Wang K, et al. Effect of microRNA-210 on prognosis and response to chemotherapeutic drugs in pediatric acute lymphoblastic leukemia. *Cancer Sci.* 2014; 105: 463-72.