

Research Paper

Prognostic Value of Lymphocyte Activation Gene-3 (LAG-3) Expression in Esophageal Squamous Cell Carcinoma

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Abstract

Esophageal squamous cell carcinoma (ESCC) is a malignant epithelial tumor with a high incidence in East Asia and the Middle East. The outcomes for ESCC patients are usually not optimal due to the recurrence and metastasis. This study aims to examine the expression and the prognostic value of LAG-3 in ESCC. We applied immunohistochemistry analysis to examine the expression of LAG-3, CD4 and CD8 in 287 ESCC cohorts. Our study demonstrated that the decreased LAG-3 expression was significantly associated with CD4 tumor-infiltrated lymphocytes (TILs) ($p=0.000$), CD8 TILs ($p=0.000$), and the advanced clinical stages ($p=0.041$) by Chi-square analysis. Kaplan-Meier survival analysis revealed that higher LAG-3 expression was positively correlated with a better overall survival (OS) ($p=0.010$) and better progression free survival (PFS) ($p=0.006$), especially in the patients at stages T1-2 status ($p=0.001$, OS; $p=0.001$, PFS), N0 status ($p=0.036$, OS; $p=0.050$, PFS), and early stages (I-II) ($p=0.006$, OS; $p=0.008$, PFS). Both high of CD4 TIL /CD8 TIL ratio and LAG-3 expression were correlated with longer OS and PFS. Cox proportional hazards regression analysis showed that LAG-3 is an independent biomarker of survival (HR, 0.724; 95% CI 0.526-0.995; $p=0.047$) ($p=0.036$). Taken together, we found that high expression of LAG-3 was correlated with an improved survival and LAG-3 is an independent predictor of survival, suggesting that LAG-3 may serve as a useful immune marker for the prognosis of ESCC.

Key words: LAG-3, ESCC, TIL, prognosis

Introduction

Esophageal carcinoma (EC) is one of the common causes of cancer-related death worldwide. There are two major subtypes of EC, which are esophageal squamous cell carcinoma (ESCC) and esophageal adenocarcinoma (EAC) [1]. ESCC is the main histological subtype of EC in East Asia and the Middle East [2]. The current treatments for ESCC are still surgery, chemotherapy and radiotherapy. Although these conventional therapies achieved certain effects, the outcomes of ESCC patients are not

significantly improved. Therefore, it is urgent to look for efficient molecular predictors and alternative therapeutic ways for ESCC.

Recently, the tumor microenvironment gains the increasing interests for developing potential anti-tumor strategies in cancer field. There were many evidences showing that the increasing number of tumor-infiltrating lymphocytes (TILs) in ESCC [3, 4]. Co-inhibitory or immune checkpoint receptors play essential roles in tumor immunity escape. Therefore,

blocking of co-inhibitory or immune checkpoint receptors might achieve potential anti-tumor immunity. For example, blockade of programmed death 1 (PD-1) expressed in exhausted T-cell has been shown a good clinical efficacy in the patients of melanoma, non-small cell lung cancer, and ovarian cancer et al [5-8].

Lymphocyte activation gene-3 (LAG-3) has been demonstrated as an important immune checkpoint in tumor immunity. LAG-3 is expressed on TILs [9], natural killer (NK) cells [9] and B cells [10]. LAG-3 have been shown to bind three ligands, major histocompatibility complex (MHC) II [11], LESTin [12] and Galectin-3 [13]. It has been reported that the expression of LAG-3 was correlated with an improved survival in both non-small-cell lung cancer (NSCLC) patients [14] and microsatellite instability-high (MSI-H) colon cancer patients [15]. Additionally, blocking LAG-3 and PD-1 induced CD8 T cells killing activation in tumors [16]. PD-1, T cell immunoglobulin-3 (TIM3), and TIGIT (T cell immunoglobulin and ITIM domain) were shown to be upregulated on TILs in ESCC patients, and PD-1 expression on TILs was correlated with late T status in this ESCC cohort [17]. However, the role of LAG-3 in ESCC still remains elusive. In this current study, we investigated the expression of LAG-3 and analyzed its prognostic values in ESCC patients.

Material and Methods

Patients

Two hundred and eighty-seven paraffin-embedded ESCC specimens were collected at Sun Yat-sen University Cancer Center. All the ESCC patients were clinically diagnosed and obtained during radical surgery prior to radiotherapy or chemotherapy from 2000 to 2006. Tissues microarrays were constructed from each specimen. The follow-up time for survivors was from 26 to 5137 days (median 1225 days). Clinical stages were defined based on the 7th edition of the Union for International Cancer Control-TNM (UICC-TNM) Classification. This study was approved by the Institutional Research Medical Ethics Committee at SYSUCC. The clinical characteristics of the ESCC patients were presented in Table 1.

Immunohistochemistry staining

The 4- μ m Formalin-fixed, paraffin-embedded ESCC tissue microarray sections were deparaffinized in xylene and with an alcohol gradient to rehydrate. The sections were immersed with three percent hydrogen peroxide to block endogenous peroxidase activity at room temperature for 10 mins, then boiled in Citrate Antigen Retrieval Solution (PH=6.0) for 5

mins in a microwaver for antigen retrieval. After that, the sections were incubated with the LAG-3 antibody (diluted at 1:150, LifeSpan BioSciences, Inc. Seattle, WA, U.S.), CD4 antibody (diluted at 1:100, ZSGB-BIO, Beijing, China), CD8 antibody (diluted at 1:100, ZSGB-BIO, Beijing, China) in a wet chamber for one and half hrs at 37°C. During the next day, the sections were washed with PBS three times and then incubated with a secondary antibody for 30 mins at 37°C. After that, the sections were stained with 3,3-diaminobenzidine for 2 mins and then counterstained with Mayer's hematoxylin to stain nucleus. At the end, the sections were dehydrated and mounted.

Table 1. Clinicopathologic characteristics of 287 patients

Clinicopathologic parameters	Cases (%)
Age	
Median (57)	
Range (32-81)	
≤57	150 (52.3)
>57	137 (47.7)
Gender	
Male	212 (73.9)
Female	75 (26.1)
T status	
T1-2	83 (28.9)
T3-4	204 (71.1)
N status	
N0	152 (53.0)
N1/2/3	135 (47.0)
M	
No	281 (97.9)
Yes	6 (2.1)
Stage	
I-II	174 (60.6)
III-IV	113 (39.4)
Grading	
G1	51 (17.8)
G2	166 (57.8)
G3	70 (24.4)
Recurrence	
No	274 (95.5)
Yes	13 (4.5)
Metastasis	
No	207 (72.1)
Yes	80 (27.9)

Evaluation of Immunohistochemistry

The immunoreactivities were scored by two independent pathologies blinded to the clinical characteristics of these ESCC patients. In the tumor region, LAG-3, CD4, and CD8 were scored as the percentage of TILs. An optional cut-off value for high and low expression was determined by the median of the score of LAG-3, CD4, and CD8, respectively. All the ESCC were divided into two subgroups according to the median score of LAG-3 immunostaining.

Statistical analysis

Statistical analysis was performed using SPSS software version 19.0. The correlation between LAG-3 and clinicopathological status of ESCC patients was examined by Spearman's rank correlation analysis and Chi-square test. Both progression-free survival (PFS) and overall survival (OS) curves for both high and low expression of LAG-3 were plotted by Kaplan-Meier analysis and log-rank test. Univariate and multivariate regression analysis were conducted using Cox proportional hazards regression model. A p -value <0.05 was considered as statistically significant.

Results

Patient characteristics

The clinical and histopathologic data for all 287 ESCC patients are described in Table 1. The median age was 57 (range from 32 to 81). Around 71.1% of ESCC patients are at late T status (T3+T4), and 47% patients have lymph node metastasis. According to the UICC guideline, there were 39.4% patients are at late clinical stages (III+IV). About 17.8%, 57.8%, and 24.4% patients were diagnosed with well differentiated (G1), intermediately differentiated (G2), poorly differentiated ESCC (G3) based on their histopathological grades. There were 85 patients were found with either local recurrence ($n=13$) or distant metastasis ($n=80$) during the follow-up time.

Association of LAG-3 expression with clinicopathological features

To determine whether LAG-3 expression was correlated with clinicopathological features, all the ESCC patients are divided into high and low LAG-3 expression groups. In our study, the cut-off score for the expression of LAG-3 was set at 0.001. If ESCC patients with negative staining intensity for LAG-3 were designated as low expression of LAG-3; if ESCC patients with visual staining intensity for LAG-3 were defined as high expression of LAG-3 (supplemental figure 1). High expression of LAG-3 was observed in 172/287 (59.9%) of ESCC patients. LAG-3 expression was significantly associated with clinical stages ($p=0.041$). Lag-3 expression was correlated with CD4 TIL ($p=0.000$) and CD8 TIL ($p=0.000$), similarly to the other reports [18, 19].

High numbers of CD4 TIL and CD8 TIL have been reported in ESCC [3, 4]. In our study, the number of CD4 TIL was negatively correlated to N status in ESCC patients (supplemental Table 1) ($p=0.044$). The number of CD8 TIL (supplemental Table 2) and the ratio of CD4 TIL/CD8 TIL (supplemental Table 3) were not associated with any clinicopathological

parameters of these ESCC patients. Furthermore, CD4 TIL/CD8 TIL is also positively correlated with LAG-3 expression ($p=0.019$, Table 2).

Table 2. Relationship between LAG-3 expression and various patient characteristics

Characteristics	No of patients	Expression of Lag3		P
		Low	High	
Patients				
Age				0.980
≤57	150	90	60	
>57	137	82	55	
Gender				0.891
Male	212	128	84	
Female	75	44	31	
T status				0.468
T1-2	83	47	36	
T3-4	204	125	79	
N status				0.142
N0	152	85	67	
N1/2/3	135	87	48	
M				0.617
No	281	169	112	
Yes	6	3	3	
Stage				0.041
I-II	174	96	78	
III-IV	113	76	37	
Recurrence				0.904
Yes	274	164	110	
No	13	8	5	
Metastasis				0.176
Yes	207	119	88	
No	80	53	27	
CD8 TIL				0.000
≤0.035	146	105	41	
>0.035	141	67	74	
CD4 TIL				0.000
≤0.01	152	109	43	
>0.01	135	63	72	
CD4 TIL/CD8 TIL				0.019
≤0.3342	173	96	76	
>0.3342	114	48	66	

Correlation of LAG-3 expression with overall survival

Among 287 ESCC patients, there were a total 172 (59.9%) cancer-related deaths at the final follow up. The five-year overall survival (OS) rate was 52.4% for this whole ESCC cohort (Figure 1A). The patients exhibiting a high LAG-3 expression presented a significantly longer OS (median, high vs low: 2202 vs 2664 days, log-rank test $p=0.010$) (Figure 1B). Further stratification of patient groups based on T, N, and clinical stages displayed that the correlation of high LAG-3 expression and higher OS was statistically significant in T1-2 status patients (median, high vs low: 2961 vs 1011 days, log-rank test $p=0.010$) (Figure 1C), N0 status patients (median, high vs low: 2703 vs 2361 days, log-rank test $p=0.038$) (Figure 1E), and clinical stages I-II patients (median, high vs low: 2244 vs 1691 days, log-rank test $p=0.006$) (Figure 1G) with ESCC.

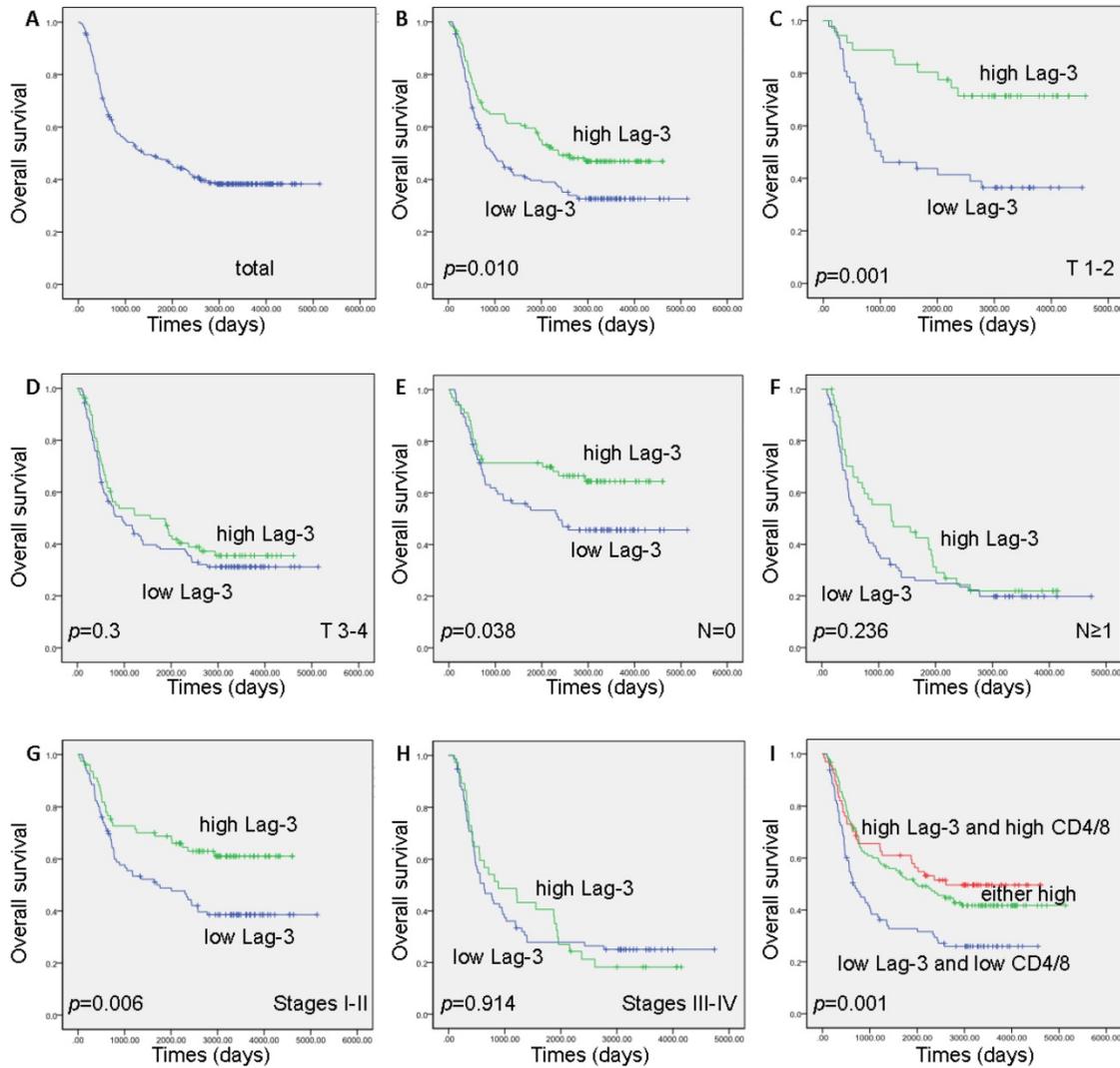


Figure 1. Kaplan-Meier survival curve and log-rank test analysis showing the association between LAG-3 expression and overall survival of ESCC patients. **A.** The five-year overall survival (OS) rate was 52.4% of 297 ESCC patient; **B.** High LAG-3 expression level was significantly correlated to OS ($p=0.010$) in all ESCC patients; **C.** High LAG-3 expression was significantly associated with OS ($p=0.001$) in ESCC patients at T 1-2 status; **D.** No significant difference in OS rate was found between LAG-3 high-expression and low-expression in ESCC patients at T 3-4 status; **E.** High LAG-3 expression was significantly associated with OS ($p=0.038$) in ESCC patients at N0 status; **F.** No significant difference in OS rate was found between LAG-3 high-expression and low-expression in ESCC patients at $N\geq 1$ status; **G.** High LAG-3 expression was significantly associated with OS ($p=0.006$) in ESCC patients at Stages I-II; **H.** No significant difference in OS rate was found between LAG-3 high-expression and low-expression in ESCC patients at Stages III-IV; **I.** Both high of CD4 TIL /CD8 TIL ratio and LAG-3 expression were correlated with longer survival ($p=0.001$).

The patients presenting high CD4 TILs seemed to have a longer OS but this correlation did not have a significant statistic difference (median, high vs low: 1942 vs 994 days, log-rank test $p=0.184$) (Supplemental Figure 2A). However, high CD8 TIL was not associated with OS (median, high vs low: 1885 vs 1225 days, log-rank test $p=0.696$) (Supplemental Figure 2B). The patients having a high CD4 TIL/CD8 TIL were correlated with a significantly improved OS (median, high vs low: 2299 vs 906 days, log-rank test $p=0.015$) (Supplemental Figure 2C). Combining of CD4 TIL /CD8 TIL ratio with LAG-3, both high of CD4 TIL /CD8 TIL ratio and LAG-3 expression were correlated with longer survival, whereas low of CD4 TIL /CD8 TIL ratio and

LAG-3 expression were correlated with shorter survival (median, high, moderate vs low: 2607, 1956 vs 649 days, log-rank test $p=0.001$) (Figure 1I).

In addition, we stratified the patients into two subgroups according to T status and stages. Interestingly, high CD8 TIL seems to have a correlation with good prognosis at T 1-2 status (supplemental figure 3A) and at early stages (supplemental figure 3C) but with a worse survival at late stages (supplemental figure 3D), suggesting that CD8 TIL exhaustion might play an important role in tumor progress at late stages. A large scale of ESCC samples or flow cytometry analysis of exhausted CD8 TIL will be further investigated.

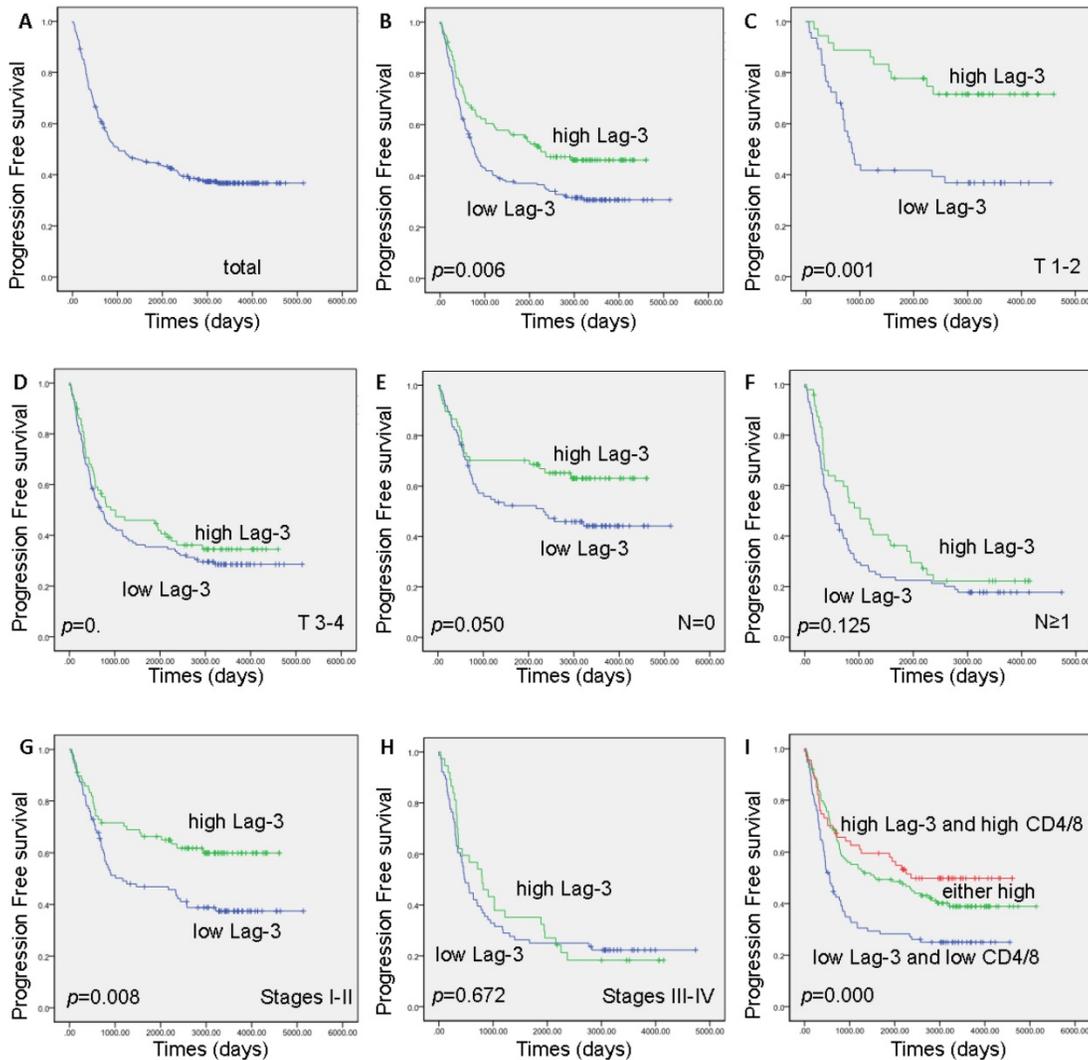


Figure 2. Kaplan-Meier survival curve and log-rank test analysis showing the association between LAG-3 expression and progression free survival of ESCC patients. **A.** The five-year progression free survival (PFS) rate was 55.3% of 297 ESCC patient; **B.** High LAG-3 expression level was significantly correlated to PFS ($p=0.006$) in all ESCC patients; **C.** High LAG-3 expression was significantly associated with PFS ($p=0.000$) in ESCC patients at T 1-2 status; **D.** No significant difference in PFS rate was found between LAG-3 high-expression and low-expression in ESCC patients at T 3-4 status; **E.** High LAG-3 expression was significantly associated with PFS ($p=0.050$) in ESCC patients at N0 status; **F.** No significant difference in PFS rate was found between LAG-3 high-expression and low-expression in ESCC patients at $N\geq 1$ status; **G.** High LAG-3 expression was significantly associated with PFS ($p=0.008$) in ESCC patients at Stages I-II; **H.** No significant difference in PFS rate was found between LAG-3 high-expression and low-expression in ESCC patients at Stages III-IV; **I.** Both high of CD4 TIL /CD8 TIL ratio and LAG-3 expression were correlated with longer survival ($p=0.000$).

Correlation of LAG-3 expression with progression-free survival

A total 177 (61.7%) cancer-related progression were found at the final follow up. The five-year progression-free survival (PFS) rate was 55.3% (Figure 2A). A high LAG-3 expression exhibited a significantly longer PFS in this ESCC cohort (2050 vs 2571 days, log-rank test $p=0.006$) (Figure 2B). Furthermore, the correlation of high LAG-3 expression and improved PFS was statistically significant in T1-2 status patients (median, high vs low: 2903 vs 859 days, log-rank test $p=0.001$) (Figure 2C), N0 status patients (median high vs low: 2787 vs 2331 days, log-rank test $p=0.050$) (Figure 2E), and clinical stages I-II patients (median, high vs low: 2592

vs 1168 days, log-rank test $p=0.000$) (Figure 2G) with ESCC.

Although the patients with higher CD4 TILs seemed to have a longer PFS, this correlation did not have a significant statistic difference (median, high vs low: 1885 vs 816 days, log-rank test $p=0.164$) (Supplemental Figure 2D). High CD8 TILs was not associated with PFS (median, high vs low: 911 vs 1027 days, log-rank test $p=0.727$) (Supplemental Figure 2E). Interestingly, a high CD4 TIL/CD8 TIL ratio was correlated with a significantly longer PFS in these ESCC patients (median, high vs low: 2171 vs 772 days, log-rank test $p=0.009$) (Supplemental Figure 2F). Moreover, both high of CD4 TIL /CD8 TIL ratio and LAG-3 expression were associated with longer PFS,

whereas low of CD4 TIL /CD8 TIL ratio and LAG-3 expression were associated with shorter PFS (median, high, moderate vs low: 2361, 1578 vs 557 days, log-rank test $p=0.000$) (Figure 2I).

The prognostic value of LAG-3 by Cox proportional hazards survival analysis

Next, we examined OS and DFS using Cox proportional hazards model to determine whether LAG-3 expression could serve as an independent biomarker. A series of factors, including age, gender, T, N, M, clinical stages, CD4 TIL/CD8 TIL, and LAG3 expression, were entered into the univariate Cox regression analysis in Table 3 to assess their impact on the OS and PFS of ESCC patients. Based on this analysis, T status, N status, clinical stages, CD4 TIL/CD8 TIL, and LAG-3 expression were found to impact OS and PFS.

The variables demonstrated to be significant in the univariate analysis were further analyzed by multivariate Cox regression analysis. The multivariate analysis demonstrated independent predictors of OS and PFS were LAG-3 expression (OS: HR, 0.724; 95% CI 0.526-0.995; $p = 0.047$; PFS: HR, 0.714; 95% CI 0.521-0.978; $p = 0.036$), T status (OS: HR, 1.625; 95% CI 1.052-2.510; $p = 0.029$; PFS: HR, 1.749; 95% CI 1.133-2.698; $p = 0.012$), and N status (OS: HR, 2.599; 95% CI 1.563-4.323; $p = 0.000$; PFS: HR, 2.704; 95% CI 1.624-4.500; $p = 0.000$)(Table 3).

Table 3. Univariate and Multivariate analysis with the COX proportional hazards for predictor of OS, and PFS of ESCC patients

Prognostic factors	Univariate			Multivariate		
	HR	CI	P	HR	CI	P
OS						
Gender	0.891	0.655-1.213	0.465
Age	0.822	0.608-1.109	0.200
T	1.720	1.203-2.459	0.003*	1.625	1.052-2.510	0.029*
N	2.409	1.771-3.278	0.000*	2.599	1.563-4.323	0.000*
M	0.902	0.288-2.826	0.860			
Stages	2.082	1.541-2.811	0.000*	0.815	0.472-1.408	0.463
CD4 TIL/CD8 TIL	0.689	0.510-0.932	0.015*	0.771	0.570-1.044	0.093
LAG-3	0.648	0.475-0.886	0.006*	0.724	0.526-0.995	0.047*
PFS						
Gender	0.926	0.688-1.247	0.614
Age	0.832	0.626-1.131	0.252
T	1.829	1.281-2.611	0.001*	1.749	1.133-2.698	0.012*
N	2.430	1.793-3.292	0.000*	2.704	1.624-4.500	0.000*
M	0.854	0.273-2.675	0.786			
Stages	2.105	1.565-2.831	0.000*	0.790	0.458-1.362	0.396
CD4 TIL/CD8 TIL	0.674	0.501-0.907	0.009*	0.761	0.564-1.026	0.073
LAG-3	0.648	0.475-0.886	0.006*	0.714	0.521-0.978	0.036*

*Statistically significant difference

Abbreviations: OS: overall survival; PFS: progression free survival; LAG-3: lymphocyte activation gene-3; HR: hazard ratio; CI: confidential interval

Discussion

Since cancer immunotherapy was named "breakthrough of the year" by Science in 2013, cancer immunology has presented a totally different way of curing cancer by targeting the host immune system instead of tumor cells [20]. Immune checkpoints are the most important immune proteins in regulating self-tolerance and escape from the immune system. Besides the broadly studies of two Immune checkpoints (cytotoxic T lymphocyte-associated molecule-4 (CTLA-4) and PD-1)[5-7, 21], more attentions have been gained in co-inhibitory receptors, including TIM3, TIGIT, and LAG-3 et al [22].

Sigurd M. Hald reported that intraepithelial-LAG-3 and stromal-LAG-3 were both associated with improved Disease-specific Survival (DSS) and OS in NSCLC [14]. Additionally, LAG-3+ TILs are an independent positive prognostic factor in stage I-IIIB NSCLC [14]. In this ESCC cohort, we demonstrated that decreased LAG-3 expression was significantly associated with advanced clinical stages (Table 2). ESCC patients with higher LAG-3 expression was positively correlated with a better OS and PFS, especially in the patients at T1-2 status, N0 status, and early stages (I-II) (Figure 1 and 2). Cox proportional hazards regression analysis showed that LAG-3 is an independent predictor of survival (Table 3). Taken together, our results suggested that LAG-3 plays an important role in the development of ESCC.

LAG-3 has been shown to be presented on CD4+, CD8+, and NK cells [9]. LAG-3 expression has been reported to correlate with other immune markers [18]. CD8, PD-1, and LAG-3 have been shown a strong correlation in breast cancer [18]. Both PD-1 and LAG-3 are demonstrated as inhibitory receptors, their expression might suggest insufficient immune response [22]. In addition, LAG-3 was mainly expressed in the CD4⁺CD25⁻ T cells, which showed high expression levels of another two inhibitory immune checkpoint protein, PD-1 and TIM3[19]. LAG-3-expressed CD4⁺CD25⁻ T cells were detected more in metastasis than in primary tumors, suggested these T cells might play important roles in cancer immune escape [19]. Here, we showed that LAG-3 expression was significantly associated with the presence of CD4 TIL and CD8 TIL, as well as the ratio of CD4/CD8 TIL (Table 2). The combination of both high LAG-3 expression and high ratio of CD4/CD8 TIL was also presented a longer OS and PFS than both low of them (Figure 1 and 2). Similarly to other reports, the increased numbers of CD4 TILs and CD8 TILs were correlated to good survival in ESCC and NSCLC et al. [4, 23]. Stromal TILs was identified as independently prognostic factor in Stage III-IVa ESCC [24]. Exhausted CD8 TILs contributes to cancer

progress [25]. Tumor-infiltrating interleukin (IL)-17A-producing cells (Th17 cells) are mainly CD4 positive and FoxP3 negative [26] in ESCC. Th17 cells were correlated with CD8 and CD1a dendritic cells (DCs) [26, 27], enhanced the cytotoxic activities of NK cells [26]. Moreover, the high level of Th17 cells was associated with good prognosis in ESCC patients [27], suggesting that Th17 cells play an important role in immune regulation during ESCC development. In our study, the higher ratio of CD4 TIL/CD8 TIL was associated with a longer survival in our cohort, indicating the dysregulation of CD4 TIL and CD8 TIL in advanced stages of ESCC. Further investigation is needed to demonstrate the percentage of LAG-3 expression on different subtypes of TILs and the clinical effects of blocking LAG-3 in ESCC.

Conclusion

In summary, we found decreased LAG-3 expression was significantly associated with advanced clinical stages. Furthermore, higher LAG-3 expression was positively correlated with a better survival and LAG-3 is an independent predictor of survival, suggesting that LAG-3 may serve as a useful immune marker for the prognosis of ESCC.

Supplementary Material

Supplementary figures and tables.

<http://www.jcancer.org/v09p4287s1.pdf>

Acknowledgements

The authenticity of this article has been validated by uploading the key raw data onto the Research Data Deposit public platform (www.researchdata.org.cn), with the approval RDD number as RDDB2018000373.

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Competing Interests

The authors have declared that no competing interest exists.

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